eISBN: 978-1-68108-233-2 ISBN: 978-1-68108-234-9 elSSN: 2213-9915 ISSN: 2451-8891

Bentham 뎢 Books

Advances in Cancer Drug Targets

Volume 3

Editor: Atta-ur-Rahman, FRS

Advances in Cancer Drug Targets *Volume 3*

Atta-ur-Rahman, FRS

Honorary Life Fellow, Kings College, University of Cambridge, UK

Advances in Cancer Drug Targets

[Volume3]

ISSN (Online): 2213-9915

ISSN (Print): 2451-8891

Advances in Cancer Drug Targets

Editor: Atta-ur-Rahman, FRS

ISBN (eBook): 978-1-68108-233-2

ISBN (Print): 978-1-68108-234-9

©[2016], Bentham eBooks imprint.

Published by Bentham Science Publishers - Sharjah, UAE. All Rights Reserved.

BENTHAM SCIENCE PUBLISHERS LTD.

End User License Agreement (for non-institutional, personal use)

This is an agreement between you and Bentham Science Publishers Ltd. Please read this License Agreement carefully before using the ebook/echapter/ejournal (**"Work"**). Your use of the Work constitutes your agreement to the terms and conditions set forth in this License Agreement. If you do not agree to these terms and conditions then you should not use the Work.

Bentham Science Publishers agrees to grant you a non-exclusive, non-transferable limited license to use the Work subject to and in accordance with the following terms and conditions. This License Agreement is for non-library, personal use only. For a library / institutional / multi user license in respect of the Work, please contact: permission@benthamscience.org.

Usage Rules:

- 1. All rights reserved: The Work is the subject of copyright and Bentham Science Publishers either owns the Work (and the copyright in it) or is licensed to distribute the Work. You shall not copy, reproduce, modify, remove, delete, augment, add to, publish, transmit, sell, resell, create derivative works from, or in any way exploit the Work or make the Work available for others to do any of the same, in any form or by any means, in whole or in part, in each case without the prior written permission of Bentham Science Publishers, unless stated otherwise in this License Agreement.
- 2. You may download a copy of the Work on one occasion to one personal computer (including tablet, laptop, desktop, or other such devices). You may make one back-up copy of the Work to avoid losing it. The following DRM (Digital Rights Management) policy may also be applicable to the Work at Bentham Science Publishers' election, acting in its sole discretion:
- 25 'copy' commands can be executed every 7 days in respect of the Work. The text selected for copying cannot extend to more than a single page. Each time a text 'copy' command is executed, irrespective of whether the text selection is made from within one page or from separate pages, it will be considered as a separate / individual 'copy' command.
- 25 pages only from the Work can be printed every 7 days.

3. The unauthorised use or distribution of copyrighted or other proprietary content is illegal and could subject you to liability for substantial money damages. You will be liable for any damage resulting from your misuse of the Work or any violation of this License Agreement, including any infringement by you of copyrights or proprietary rights.

Disclaimer:

Bentham Science Publishers does not guarantee that the information in the Work is error-free, or warrant that it will meet your requirements or that access to the Work will be uninterrupted or error-free. The Work is provided "as is" without warranty of any kind, either express or implied or statutory, including, without limitation, implied warranties of merchantability and fitness for a particular purpose. The entire risk as to the results and performance of the Work is assumed by you. No responsibility is assumed by Bentham Science Publishers, its staff, editors and/or authors for any injury and/or damage to persons or property as a matter of products liability, negligence or otherwise, or from any use or operation of any methods, products instruction, advertisements or ideas contained in the Work.

Limitation of Liability:

In no event will Bentham Science Publishers, its staff, editors and/or authors, be liable for any damages, including, without limitation, special, incidental and/or consequential damages and/or damages for lost data and/or profits arising out of (whether directly or indirectly) the use or inability to use the Work. The entire liability of Bentham Science Publishers shall be limited to the amount actually paid by you for the Work.

General:

- 1. Any dispute or claim arising out of or in connection with this License Agreement or the Work (including non-contractual disputes or claims) will be governed by and construed in accordance with the laws of the U.A.E. as applied in the Emirate of Dubai. Each party agrees that the courts of the Emirate of Dubai shall have exclusive jurisdiction to settle any dispute or claim arising out of or in connection with this License Agreement or the Work (including non-contractual disputes or claims).
- 2. Your rights under this License Agreement will automatically terminate without notice and without the need for a court order if at any point you breach any terms of this License Agreement. In no event will any delay or failure by Bentham Science Publishers in enforcing your compliance with this License Agreement constitute a waiver of any of its rights.
- 3. You acknowledge that you have read this License Agreement, and agree to be bound by its terms and conditions. To the extent that any other terms and conditions presented on any website of Bentham Science Publishers conflict with, or are inconsistent with, the terms and conditions set out in this License Agreement, you acknowledge that the terms and conditions set out in this License Agreement shall prevail.

Bentham Science Publishers Ltd. Executive Suite Y - 2 PO Box 7917, Saif Zone Sharjah, U.A.E. Email: subscriptions@benthamscience.org



CONTENTS

PREFACE				
			1. INTRODUCTION: NEUTROPHIL ELASTASE/A-1ANTITRYPSIN IMBALANCE	AS A LINK
			BETWEEN CHRONIC OBSTRUCTIVE PULMONARY DISEASE AND LUNG CANC	ER 4
2. MULTIFACETED FUNCTIONS OF NEUTROPHIL ELASTASE IN LUNG CANCER	7			
3. ENDEGNENOUS NEUTROPHIL ELASTASE INHIBITORS				
3.1. Proteinaceous Inhibitors				
3.2. Natural Compounds	12			
3.2.1. Glycosaminoglycans				
3.2.2. Phenolics	13			
3.2.3. Triterpenoids				
3.2.4. Fatty Acids and Peptide Derivatives	19			
4. DESIGN OF DUAL NEUTROPHIL ELASTASE / MMP INHIBITORS				
DESIGN OF DUAL HNE-MMP INHIBITORS				
CONCLUDING REMARKS				
ADDITIONAL MATERIAL				
1. Molecular Modeling and Molecular Graphics				
2. Ligand and Receptor Preparation				
3. Docking Protocol				
CONFLICT OF INTEREST				
DISCLOSURE				
ACKNOWLEDGEMENTS				
ABBREVIATIONS				
REFERENCES				
CHAPTER 2 INHIBITION OF MEMBRANE COMPLEMENT INHIBITOR EXPRESSION ((CD46, CD55,			
D57) BI SIRNA SENSITIZES TUMOR CELLS TO COMPLEMENT ATTACK				
INTRODUCTION				
Call Culture				
SIRNA Sequences				
SIKNA Iransiecuon				
Flow Cytometry				
Comprement-methaded Cytotoxicity Assay (CDC)				
Deal Time DT DCD				
NGAI-IIIICNI-PUN				
Statistical Allalysis				
Design of siDNAs Specific for CD46 CD55 and CD50				
Sign of signated Decempendation of mCDP E				
SIKINA-Inediated Downregulation of muckP Expression				
SIGNA-inclusion Augmentation of Lumor Cell Complement Lysis and Opsonization				
Thic-coulst of SixivA-Inducto inc.r finite on U-imit - DNA E-marchine Martin				
Stable Downregulation of CD39 Using an Hairpin SiKINA Expression vector				
DISCUSSION				

CONFLICT OF INTEREST	71
DISCLOSURE	
ACKNOWLEDGEMENT	
ABBREVIATIONS	
REFERENCES	
CHAPTER 3 POINTS OF THERAPEUTIC INTERVENTION ALONG THE WNT SIG	SNALING
PATHWAY IN HEPATOCELLULAR CARCINOMA	
INTRODUCTION	
THE WNT SIGNALING PATHWAY	80
Overview of the Wnt Signaling	80
The Wnt/β-catenin Pathway	82
Aberrant Activation of the Wnt/β-catenin Pathway in HCC	83
TARGETING THE WNT/B-CATENIN PATHWAY IN HCC	
Targeting the Upstream Components	
Endogenous Inhibitors of the Ligand/Receptor Complex	91
Targeting Wnt Ligands and FZD Receptors	
Targeting the Dishevelled Protein	93
Cellular Trafficking and Targets	
Targeting the β-catenin Destruction Complex	
Targeting the β-catenin/TCF Transcriptional Complex	
Pitfalls in Targeting the Wnt/β-catenin Pathway	
CONCLUSIONS AND PERSPECTIVES	
CONFLICT OF INTEREST	100
DISCLOSURE	100
ACKNOWLEDGMENT	100
ABBREVIATIONS	101
REFERENCES	102
CHARTER 4 COLLABORATION OF EDITHELIAL MECENCHAMAL TRANSITION AND	CANCED
CHAFTER 4 COLLABORATION OF EFFITIELIAL MESENCHYMAL TRANSITION AND	UANUEK ICED
STEM CELLS: SINISTER ROUTES FOR CHEMORESISTANT RECURRENT OVARIAN CAP	ICER
	119
PATHOLOGY OF OVARIAN CANCER	119
IRANSITION FROM EPITHELIAL TO MESENCHYMAL PHENOTYPE AND THE PROG	KESSION 100
UF CANCER	122
EVIDENCE OF ENIT IN OVARIAN CANCER	123
STEM CELLS IN NORMAL OVARIES AND OVARIAN CANCER	123
ASSOCIATION OF EMT AND CSCs. A MEDGED FOR DOTENTIAL CHEMODESIST	129
ASSOCIATION OF EMITAND CSCS: A MERGER FOR FOTENTIAL CHEMORESIST	ANCE IN 120
Cigniatin Induced EMT Concretes Quarian Cancer Stam Like Colle: A Study on the OVCA 422 (Coll Line of
an Experimental Model	132
NFW THERAPEUTIC APPROACHES AND CONCLUSION	132
CONFLICT OF INTEREST	1/1
DISCLOSURF	142
ACKNOWI FDGFMFNTS	1/12
ABREVIATIONS	142
REFERENCES	143

CHAPTER 5 OXALIPLATIN-MEDIATED INHIBITION OF SURVIVIN INCREASES SENSITIVITY OF

-	
INTRODUCTION	
MATERIALS AND METHODS	
Cell Culture	
Plasmid and Antisense Transfection Efficiency	
Drug Treatment	
MTT (methylthiazolyldiphenyl-tetrazolium bromide) Assay	
Western Blot Analysis	
Real-Time PCR	
TUNEL (Terminal Deoxyribonucleotidyl Transferase Mediated dUTP Nick End Labeling) / Statistical Analysis	Assay
RESULTS	
Detection of Survivin Gene Expression	
Effect of Paclitaxel and Oxaliplatin on Cell Growth and Survival	
Survivin Expression Reduces Paclitaxel-mediated Cytotoxicity	
Oxaliplatin Sensitizes Cancer Cells to Paclitaxel by Inhibiting Survivin	
Inhibition of Survivin by a siRNA Method Induces Paclitaxel-mediated Cell Death	
DISCUSSION	
CONFLICT OF INTEREST	
DISCLOSURE	
ACKNOWLEDGEMENTS	
ABBREVIATIONS	
	•••••
REFERENCES HAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS	YTUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions	YTUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction	YTUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMARY GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMARY GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures	YTUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments	YTUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments Measurement of Steroid Sulfatase Activity	YTUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase	YTUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS Histopathology of Mammary Tumors	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS Histopathology of Mammary Tumors Evolution of Body Weight	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS Histopathology of Mammary Tumors Evolution of Body Weight Tumor Growth	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS Histopathology of Mammary Tumors Evolution of Body Weight Tumor Growth Survival Probability	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS Histopathology of Mammary Tumors Evolution of Body Weight Tumor Growth Survival Probability Serum Estradiol Concentration and Uterine Weight	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS Histopathology of Mammary Tumors Evolution of Body Weight Tumor Growth Survival Probability Serum Estradiol Concentration and Uterine Weight Survival Probability	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS Histopathology of Mammary Tumors Evolution of Body Weight Tumor Growth Survival Probability Serum Estradiol Concentration and Uterine Weight Sulfatase Activity and Expression	Y TUMORS
REFERENCES IAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS Histopathology of Mammary Tumors Evolution of Body Weight Tumor Growth Survival Probability Serum Estradiol Concentration and Uterine Weight Sulfatase Activity and Expression DISCUSSION CONFLICT OF INTEREST	Y TUMORS
REFERENCES HAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' GULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS Histopathology of Mammary Tumors Evolution of Body Weight Tumor Growth Survival Probability Serum Estradiol Concentration and Uterine Weight Sulfatase Activity and Expression DISCUSSION CONFLICT OF INTEREST DISCLOSURE	Y TUMORS
REFERENCES HAPTER 6 MELATONIN INHIBITS THE GROWTH OF DMBA-INDUCED MAMMAR' COULATING ESTROGEN SULFATASE ENZYME INTRODUCTION MATERIAL AND METHODS Animals and Housing Conditions Tumor Induction Experimental Design Surgical Treatments, Tumor Size and Number, Survival Rates and Autopsy Procedures Estrone Sulfate and Melatonin Treatments Measurement of Steroid Sulfatase Activity Measurement of mRNA Expression of Sulfatase Statistical Analysis RESULTS Histopathology of Mammary Tumors Evolution of Body Weight Tumor Growth Survival Probability Serum Estradiol Concentration and Uterine Weight Sulfatase Activity and Expression DISCUSSION CONFLICT OF INTEREST DISCLOSURE ACKNOWLEDGEMENTS	Y TUMORS

 CHAPTER 7
 ROLE OF mTOR SIGNALING IN TUMOR CELL MOTILITY, INVASION AND

 METASTASIS
 207

RODUCTION	208
mTOR Strcture and Signaling Complexes	209
mTOR Signaling Complexes	210
mTORC1	210
mTORC2	214
mTOR Inhibitors	215
The Role of mTOR Signaling Pathway in Cell Motility and Invasion	218
mTORC1 Signaling in Cell Motility and Invasion	218
mTORC2 Signaling in Cell Motility and Invasion	223
Rapamycin and Tumor Metastases	225
[MARY	227
IFLICT OF INTEREST	228
CLOSURE	228
NOWLEDGMENTS	228
ERENCES	228
ER 8 STRUCTURE-ACTIVITY STUDIES ON ARYLAMIDES AND ARYSULFONA	AIDES RAS
ORS	245
RODUCTION	245
ULTS AND DISCUSSION	248
Chemistry	248
Biochemistry	249
Interaction between Compound 1 (SCH-53870) and Ras: Preliminary Observations	249
Nucleotide Exchange Inhibition Experiments with Compounds 1-17	252
Effect of Compounds 1-4 on Mammalian Cell Growth	254
IPUTATIONAL ANALYSIS	255
Docking Calculations	255
ICLUSION	257
FERIALS AND METHODS	259
Expression and Isolation of Proteins	259
Measurement of GEF-Stimulated Guanine Nucleotide Exchange on p21 h-Ras	259
Measurement of Dissociation Rate	259
Two-Hybrid System	
Cell Cultures and Growth Conditions	
Docking Calculations	
PLEMENTARY MATERIAL	261
FLICT OF INTEREST	
CLOSURE	261
NOWLEDGMENTS	
REVIATIONS	
ERENCES	262
ERENCES	

PREFACE

The 3rd volume of the book Series "*Advances in Cancer Drug Targets*" comprises eight chapters written by the leading experts in this field. It is an outstanding collection of well written chapters on cancer drug targets in the field of pharmacology, molecular biology and biochemistry.

Human neutrophil elastase (HNE) plays an important role in the development of chronic obstructive pulmonary diseases. In chapter 1, Alix *et al.*, explain its involvement in non-small cell lung cancer progression. Natural compounds and/or synthesized agents which antagonize HNE activity have been comprehensively reviewed in this chapter. They also focus on substances (*i.e.* lipids and derivatives, phenolics) that exhibit an inhibitory bifunctionality towards HNE and matrix metalloproteinases (MMPs), particularly MMP-2.

The efficacy of cancer-immunotherapy with complement-activating monoclonal antibodies is restricted by over-expression of one or more membrane-bound complement regulatory proteins (mCRPs: CD46, CD55, CD59) that are present on the surface of neoplastic cells. Kirschfink *et al.*, in chapter 2 discuss small interfering RNAs (siRNAs) for post-transcriptional gene knock down of CD46, CD55 and CD59 aiming to sensitize tumor cells.

Hepatocellular carcinoma (HCC) is the third most common cause of deaths from cancer worldwide. There is growing evidence that the deregulation of Wnt/ β -catenin signaling pathway plays a critical role in hepatic oncogenesis and mainly occurs at the early stage of hepatocarcinogenesis. In chapter 3, Kim and Wands have summarized the potential molecular targets related to the Wnt/ β -catenin signaling pathway along with their therapeutic applications.

A major challenge in treating ovarian cancer is to overcome intrinsic and acquired. Chapter 4 by Ahmed *et al.* presents the recent advances in our understanding of the cellular origin and the molecular mechanisms defining the basis of cancer initiation and malignant transformation with respect to epithelial-mesenchymal transition (EMT) of ovarian cancer cells.

Due to the high expression of Survivin in various carcinomas, it is one of the key antiapoptotic proteins. It is also associated with their biologically aggressive characteristics and drug resistance. Bisen *et al.*, in chapter 5 elaborate the efficacy of combination of oxaliplatin and paclitaxel as a potential strategy in controlling HNSCC cell proliferation. This review highlights the fact that the co-treatment of cells with paclitaxel and oxaliplatin results in a significantly higher cytotoxicity as compared to individual single drug treatment. Melatonin has oncostatic effects on different neoplasias, particularly on estrogen-dependent breast cancer. The compound acts by interacting with estrogen-responsive pathways, thus behaving as an antiestrogenic hormone. In chapter 6 by Cos *et al.*, evidence is presented that that melatonin could exert its antitumoral effects on hormone-dependent mammary tumors by down-regulating the sulfatase pathway of the tumoral tissue.

Recent studies have thrown light on the role of mammalian target of rapamycin (mTOR) in the regulation of tumor cell motility, invasion and cancer metastasis. Zhou and Huang in chapter 7 discuss the mTOR complexes and the role of mTOR signaling in tumor cell migration and invasion. The chapter also highlights the findings about the mechanism by which rapamycin inhibits cell migration, invasion and cancer metastasis.

It has been hypothesised that a phenyl hydroxylamine group linked to a second aromatic moiety generates a pharmacophore which can interact with Ras and inhibit its activation In chapter 8, Peri *et al.*, present reports on the synthesis of a library of small molecules with arylamides and arylsulfonamides groups. They also explain their biological activity to inhibit nucleotide exchange on human Ras.

I hope that the current book volume, which provides insights into the development of new approaches to anti-cancer therapy for interested researchers and pharmaceutical scientists, will be received with the same enthusiasm as the previous volumes of this Series. I am grateful to the valuable contributions made by the authors. I greatly appreciate the assistance from the editorial staff, particularly Mr. Mahmood Alam (Director Publications) and Mr. Shehzad Naqvi (Senior Manager) for their hard work and determined efforts.

Atta-ur-Rahman Kings College University of Cambridge Cambridge UK

ii

List of Contributors

Alain J.P. Alix	Institut de Chimie Moléculaire de Reims & Laboratoire de Spectroscopies et Structures Biomoléculaires, Faculté des Sciences, IFR53 Biomolécules, BP 1039, 51687, Université de Reims Champagne-Ardenne, CNRS UMR 6229, Reims Cedex, France
Alessandro Palmioli	Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy
Alicia Gonzlez	Department of Physiology and Pharmacology, School of Medicine, University of Cantabria and Instituto de Investigación Valdecilla (IDIVAL), 39011 Santander, Spain
E. Chan	Department of Obstetrics and Gynaecology, University of Melbourne, Melbourne, Australia
Carolina Martínez- Campa	Department of Physiology and Pharmacology, School of Medicine, University of Cantabria and Instituto de Investigación Valdecilla (IDIVAL), 39011 Santander, Spain
Cristina Airoldi	Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy
Enzo Martegani	Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy
Erika BOURGUET	Institut de Chimie Moléculaire de Reims, Faculté de Pharmacie, IFR53 Biomolécules, 51 rue de Cognacq-Jay, Université de Reims Champagne- Ardenne, CNRS UMR 6229, 51096 Reims Cedex, France
Francesco Peri	Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy
G. Kannourakis	Fiona Elsey Cancer Research Institute, Ballarat, Australia Federation University, , Ballarat, Australia
G.B.K.S. Prasad	School of Studies in Biochemistry, Jiwaji University, Gwalior 474011, India
Giese Thomas	Institute of Immunology, University of Heidelberg, Heidelberg, Germany
Hongyu Zhou	Department of Biochemistry and Molecular Biology, Louisiana State University, Health Sciences Center, 1501 Kings Highway, Shreveport, LA 71130-3932, USA
J.K. Findlay	Department of Obstetrics and Gynaecology, University of Melbourne, Melbourne, Australia Hudson Institute of Medical Research, Melbourne, Australia
Jack R. Wands	Institute of Immunology, Rhode Island Hospital and the Warren Alpert Medical School of Brown University, Providence, RI, USA

Janos SAPI	Institut de Chimie Moléculaire de Reims & Laboratoire de Spectroscopies et Structures Biomoléculaires, Faculté des Sciences, IFR53 Biomolécules, BP 1039, 51687, Université de Reims Champagne-Ardenne, CNRS UMR 6229, Reims Cedex, France
K. Abubaker	Department of Surgery, University of Melbourne, Melbourne, Australia
Luca De Gioia	Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy
Michael Kirschfink	Institute of Immunology, University of Heidelberg, Heidelberg, Germany
Miran Kim	Institute of Immunology, Rhode Island Hospital and the Warren Alpert Medical School of Brown University, Providence, RI, USA
Moroy Gautier	Molécules thérapeutiques in silico, Université Paris Diderot, INSERM UMR- S-973, Bât
N. Ahmed	Fiona Elsey Cancer Research Institute, Ballarat, Australia Department of Surgery, University of Melbourne, Melbourne, Australia Department of Obstetrics and Gynaecology, University of Melbourne, Melbourne, Australia Hudson Institute of Medical Research, Melbourne, Australia
N. Khan	Division of Plant-Microbe Interactions, National Botanical Research Institute, Rana Pratap Marg, Lucknow, 226 001, India
Nicolas Geis	Institute of Immunology, University of Heidelberg, Heidelberg, Germany
P. S. Bisen	School of Studies in Biotechnology, Jiwaji University, Gwalior 474011, India
Renate Rutz	Institute of Immunology, University of Heidelberg, Heidelberg, Germany
Renata Tisi	Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy
Samuel Cos	Department of Physiology and Pharmacology, School of Medicine, University of Cantabria and Instituto de Investigación Valdecilla (IDIVAL), 39011 Santander, Spain
Sandro Olivieri	Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy
Shile Huang	Department of Biochemistry and Molecular Biology, Louisiana State University, Health Sciences Center, 1501 Kings Highway, Shreveport, LA 71130-3932, USA Feist-Weiller Cancer Center, Louisiana State University, Health Sciences Center, 1501 Kings Highway, Shreveport, LA 71130-3932, USA
Simon Höne	Institute of Immunology, University of Heidelberg, Heidelberg, Germany
Sonia Colombo	Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy

iv

	ν
Sonia Fantinato	Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy
Srinivas Mamidi	Institute of Immunology, University of Heidelberg, Heidelberg, Germany
Stefanie Zell	Institute of Immunology, University of Heidelberg, Heidelberg, Germany
Stefan Schultz	Institute of Immunology, University of Heidelberg, Heidelberg, Germany
Virginia álvarez-García	Department of Physiology and Pharmacology, School of Medicine, University of Cantabria and Instituto de Investigación Valdecilla (IDIVAL), 39011 Santander, Spain
Wenhan Li	Institute of Immunology, University of Heidelberg, Heidelberg, Germany
William Hornebeck	Institut de Chimie Moléculaire de Reims, Faculté de Pharmacie, IFR53 Biomolécules, 51 rue de Cognacq-Jay, Université de Reims Champagne- Ardenne, CNRS UMR 6229, 51096 Reims Cedex, France
Z. Khan	Department of Biomedical Sciences and Pathology and Laboratory Medicine, Cedars-Sinai Medical Center, Los Angeles, CA 90048, USA

Neutrophil Elastase as a Target in Lung Cancer: the State of the Art

Gautier Moroy¹, Alain J.P. Alix^{2,*}, Janos Sapi³, William Hornebeck⁴, Erika Bourguet³

¹ Université Paris Diderot, Sorbonne Paris Cité, Molécules Thérapeutiques In Silico, Inserm UMR-S 973, 35 rue Hélène Brion, 75013 Paris, France

² Université de Reims Champagne-Ardenne, CNRS UMR 7312, Institut de Chimie Moléculaire de Reims & Laboratoire de Spectroscopies et Structures Biomoléculaires, Faculté des Sciences, SFR CAP-Santé BP 1039, 51687 Reims Cedex2, France

³ Université de Reims Champagne-Ardenne, CNRS UMR 7312, Institut de Chimie Moléculaire de Reims, Faculté de Pharmacie, SFR CAP-Santé 51 rue de Cognacq-Jay, 51096 Reims Cedex, France

⁴ Université de Reims Champagne-Ardenne, CNRS UMR 6237, Laboratoire de Biochimie Médicale, Faculté de Médecine, SFR CAP-Santé 51 rue de Cognacq-Jay, 51096 Reims Cedex, France

Abstract: Human neutrophil elastase (HNE), a main factor in the development of chronic obstructive pulmonary diseases, has been recently involved in non-small cell lung cancer progression. It can act at several levels (i) intracellularly, cleaving for instance the adaptor molecule insulin receptor substrate-1 (IRS-1) (ii) at the cell surface, hydrolyzing receptors as CD40 (iii) in the extracellular space, generating elastin fragments *i.e.* morphoelastokines which potently stimulate cancer cell invasiveness and angiogenesis.

Since decades, researchers identified natural compounds and/or synthesized agents which antagonize HNE activity that will be described in this review article. Some of these compounds might be of value as therapeutic agents in lung cancer.

Atta-ur-Rahman (Ed) All rights reserved-© 2016 Bentham Science Publishers

^{*} Address correspondence to Alain J.P. Alix: Laboratoire de Spectroscopies et Structures Biomoléculaires & Institut de Chimie Moléculaire de Reims, CNRS UMR 7312, Faculté des Sciences Exactes et Naturelles, BP 1039, 51687 Reims Cedex2, France; Tel.: + 33 6 73 87 93 43; Fax : + 33 3 26 91 80 29; Email: alain.alix@univ-reims.fr.

4 Advances in Cancer Drug Targets, Vol. 3

However, it is now widely accepted that lung tumor invasion and metastasis involve proteolytic cascades. Accordingly, we will here mainly focus our attention to natural substances able to display a dual inhibitory capacity (*i.e.* lipids and derivatives, phenolics) towards HNE and matrix metalloproteinases (MMPs), particularly MMP-2. To that purpose, we had synthesized substances named "LipoGalardin" exhibiting such inhibitory bifunctionality. At last, we will propose an original synthetic scheme for designing a potent biheaded HNE/MMP-2 inhibitor.

Keywords: Bifunctionality, Caffeic acid phenethyl ester (CAPE), (Dual) inhibitors, Elafin, Elastokines, (-)-Epigallocatechin-3-gallate (EGCG), Flexible docking, Lipogalardin, Lung cancer, Molecular modelling, Neutrophil Elastase, (Potent) angiogenic molecules, Potent biheaded lnhibitor, (Potent) chemiotactic activity.

1. INTRODUCTION: NEUTROPHIL ELASTASE/A-1ANTITRYPSIN IMBALANCE AS A LINK BETWEEN CHRONIC OBSTRUCTIVE PULMONARY DISEASE AND LUNG CANCER

Cigarette smoke is widely considered as the causative agent in chronic obstructive pulmonary disease (COPD) and lung cancer, two leading causes of death worldwide [1, 2]. This common initiating agent is able to generate in the lung reactive oxygen species resulting in NF-kappa B activation and inflammation [3] but consequences appear distinct in the two diseases. COPD is characterized mainly by matrix degradation, incomplete tissue repair with excessive apoptosis and impaired neovessel formation, while excessive DNA damage and its incomplete repair are hallmarks of lung cancer [2]. However, recent investigations pinpointed that smokers who suffer from COPD appear to be at increased risk for developing adenocarcinoma of the lung, suggesting that it might exist a link between these pathologies [4 - 6]. Such link between COPD and lung cancer has been clearly evidenced in population based study [7].

Alpha-1 antitrypsin (α -1AT) deficiency caused by the homozygous ZZ allele is responsible for liver disease in children and emphysema in young adults [8, 9]. These individuals have a shorter life span than the general population and lung cancer, an age-related disease, was not detected in patients. However, a 14 fold increased risk of lung cancer was evidenced in non-smokers α -1AT deficiency

Targeted Elastase in Lung Cancer

Advances in Cancer Drug Targets, Vol. 3 5

gene-carriers younger than 60 years of age [10]. Particularly the frequency of M1 allele and PiM1 a-1AT homozygotes in lung cancer patients was found to be significantly elevated as compared to healthy individuals [11, 12]. Alpha-1 Antitrypsin belongs to the serpin family, and it displays the highest affinity among serine proteinases for neutrophil elastase (NE) [13]. To that respect, since decades, the elastase/ α -1 antitrypsin imbalance was considered as one pivotal mechanism in the formation of emphysematous lesions characterized by intense elastin fragmentation and airways enlargement [14]. Imbalance between enzyme and inhibitor might originate from genetic deficiency of α -1AT; alternatively, the oxidation of Met 358 in the α -1AT active site markedly impairs its interactions with NE. Also, Matrix Metalloproteinases (MMP), mainly matrilysin, as overexpressed in COPD and lung cancer, can further inactivate native or oxidized α -1AT by proteolysis [15]. Altogether, these mechanisms led to the generation of excess NE in pathological lung tissues. In the search of pathological biomarkers a recent in vivo study pointed out that A α -Val360, an HNE specific fibrinogen degradation product may represent an ideal indicator for COPD disease severity and progression. This report is considered as the first in vivo data supporting physiopathological role of HNE in COPD [16]. Human NE (HNE: EC 3.4.21.37) gene encodes a 267 amino acid residues preprotein and its transcription is restricted to the promyelocytic stage of granulocyte development. Mature HNE contains 218 amino acid residues and two sites of N-glycosylation have been identified, but isoenzymes display similar kinetic constants with substrate and inhibitors. Enzyme at high concentration: 3 pg/cell is sequestered within azurophilic granules of polymorphonuclear neutrophils (PMN) and its activity is mainly controlled by compartmentalization [17, 18]. The main physiological function of NE, together with reactive oxygen species and other PMN proteases, consists in fighting against microbial action [13, 17]. Activation of PMN with bacterial products and cytokines only minimally influences NE release. On the contrary, frustrated phagocytosis or PMN necrosis as it occurred in COPD, leads to intense release of elastase from cells [17].

A recent mass spectrometric proteome analysis has evidenced that a histone H2A specific protease (H2Asp), discovered more than 35 years ago and implicated in the truncation of the histone H2A C-tail at V114 in myeloid cells is finally

CHAPTER 2

Inhibition of Membrane Complement Inhibitor Expression (CD46, CD55, CD59) by siRNA Sensitizes Tumor Cells to Complement Attack

Srinivas Mamidi, Simon Höne, Nicolas Geis, Stefanie Zell, Renate Rutz, Wenhan Li, Thomas Giese, Stefan Schultz, Michael Kirschfink^{*}

Institute of Immunology, University of Heidelberg, Heidelberg, Germany

Abstract: The efficacy of cancer-immunotherapy with complement-activating monoclonal antibodies is limited by over-expression of one or more membrane-bound complement regulatory proteins (mCRPs: CD46, CD55, CD59) on the surface of neoplastic cells.

In this study we designed small interfering RNAs (siRNAs) for posttranscriptional gene knock down of CD46, CD55 and CD59 aiming to sensitize tumor cells to complement attack and thereby exploiting complement for tumor cell destruction. Tumor cell lines of different origin, such as Du145 (prostate), BT474 (breast) and K562 (erythroleukemia) were selected for the study. FACS-analysis demonstrated that siRNA anti-CD46 (301) reduced CD46 protein expression up to 80%, siRNA anti-CD55 (255) diminished CD55 protein expression up to 49%, and CD59 protein expression was inhibited up to 82% by siRNA anti-CD59 (1339). Time course experiments revealed a long-lasting silencing effect with >50% complement regulator inhibition up to day 13. Upon mCRP knock down, complement-dependent cytotoxicity (CDC) was augmented by 20-30% for CD46, by up to 24% for CD55 and by up to 55% for CD59. The combined inhibition of all three inhibitors further enhanced CDC by up to 66%. Dependent on the cell line, CD46 and CD55 downregulation increased significantly C3 opsonization, which is known to support cell-mediated defense mechanisms. mCRP blocking antibodies were only partly able to further augment the tumor cells' susceptibility to complement lysis.

Atta-ur-Rahman (Ed) All rights reserved-© 2016 Bentham Science Publishers

50

^{*} Address correspondence to Michael Kirschfink: Institute of Immunology, University of Heidelberg, Germany, Im Neuenheimer Feld 305, 69120 Heidelberg, Germany; Tel: +49 6221 56 4076/4026; Fax: +49 6221 56 5586; Email: michael.kirschfink@urz.uni-heidelberg.de.

Inhibition of Membrane Complement

Thus, siRNA-induced inhibition of complement regulator expression clearly sensitizes malignant cells to complement attack and, if specifically targeted to the tumor, appears suited as adjuvant to improve antibody-based cancer immunotherapy.

Keywords: Complement resistance, Membrane complement regulatory proteins, siRNA, Tumor immunology.

INTRODUCTION

The potential of complement activation in the control of neoplastic cells is hampered by various protective mechanisms employed by the tumor cell. The same mechanisms, which protect normal cells from accidental complement lysis, are also involved in tumor cell resistance to complement. Among these mechanisms, the expression of the membrane-bound complement regulatory proteins (mCRP) CD46, CD55 and CD59 appears most important and consequently gained most attention [1 - 5].

Compared to the corresponding unaffected tissue, cancer cells often over-express one or more of these mCRPs. As previously suggested this may be due to selective forces caused by repeated complement attacks during neoplastic transformation [1]. Furthermore, an increasing number of studies showed that mCRP over-expression is a result of different mechanisms related to activation of tumor cells. For example, Prostaglandin E_2 can up regulate CD55 expression [6]. CD46 mRNA expression is induced by activation of the signal transducers and activators of transcription 3 (STAT-3) pathway [7], which is - in contrast to normal tissues or cells - persistently activated in most cancer cells and primary tumor tissues [8]. CD59 can be upregulated as a result of increased acetylation of p53, a process that particularly occurs in tumors [9]. It has been demonstrated that enhanced expression of membrane regulators is associated with reduced survival in cancer patients [10, 11].

The inhibitors CD46 [12] and CD55 [13] block the complement cascade at the C3 activation stage, thereby not only preventing direct complement-dependent cytotoxicity, but also limiting the generation of the anaphylatoxins C3a and C5a. Subsequently, an attenuated inflammatory reaction in combination with a reduced amount of C3 opsonins deposited on the cell surface restricts cell-mediated

52 Advances in Cancer Drug Targets, Vol. 3

Mamidi et al.

immune defense mechanisms.

CD59 [14] interferes with the assembly of the membrane attack complex (MAC) of complement *via* binding to C9, thereby preventing the formation of the lytic pore. CD59 is considered to be the most effective surface inhibitor, acting additively or even synergistically with CD46 and CD55 [15].

Neutralization of one or more membrane regulators renders tumor cells more susceptible to complement attack. This has been verified in the past by either blocking antibodies against mCRPs [1, 4, 16], enzymatic removal of the GPI-anchored CD55 and CD59 by PI-phospholipase C [17] or by cytokine-mediated downregulation [18].

We have identified CD46- and CD55-specific phosphorothioate antisenseoligonucleotides (S-ODNs) to immediately interfere with synthesis of both mCRPs in neoplastic cells leading to a significantly reduced complement resistance [19].

RNA interference (RNAi) based on small interfering RNAs (siRNAs) is considered to be more efficient than traditional antisense strategies [20]. SiRNAs are short double-stranded RNA oligomers consisting of 21-23 bp, which induce highly sequence-specific degradation of homologous mRNA, thereby silencing the target gene by posttranscriptional gene knock down [21, 22].

We here demonstrate a significant reduction of tumor cell complement resistance upon the application of siRNAs targeting all three mCRPs (CD46, CD55 and CD59).

MATERIALS AND METHODS

Cell Culture

DU145 prostate carcinoma cells ATCC (American Type Culture Collection, Manassas, VA, USA) number: HTB-81, BT474 breast carcinoma cells (ATCC number: HTB-20) and K562 erythroleukemic cells (ATCC number: CCL 243) were all cultured in RPMI 1640 (Cambrex, Verviers, Belgium), supplemented with 10% (v/v) heat-inactivated fetal calf serum (FCS, Invitrogen, Karlsruhe,

CHAPTER 3

Points of Therapeutic Intervention along the Wnt Signaling Pathway in Hepatocellular Carcinoma

Miran Kim^{*}, Jack R. Wands

Liver Research Center, Rhode Island Hospital and the Warren Alpert Medical School of Brown University, Providence, RI, USA

Abstract: Hepatocellular carcinoma (HCC) is the third most common cause related to cancer mortality worldwide. Due to frequently late diagnosis, overall prognosis of patients with liver cancer is poor. Unfortunately, there is no targeted therapeutics for the treatment of HCC except sorafenib, which has exhibited notable results in certain advanced HCC. Increasing evidences indicate that deregulation of Wnt/ β -catenin signaling pathway plays a critical role in hepatic oncogenesis and mainly occurs at the early stage of hepatocarcinogenesis. In addition, aberrant activation of the Wnt/ β -catenin signaling pathway has been linked with more aggressive HCCs. The major mechanism for aberrant activation of the signaling in HCC is caused by genetic mutations and/or altered expression of upstream components of the Wnt/ β -catenin signaling. This leads to abnormal expression of the β -catenin/TCF-responsive target genes, which regulate cell growth, apoptosis, cell motility, and invasion. Thus, intervention of the Wnt/ β -catenin signaling activity can be potential therapeutics for HCC. This review will discuss the identified potential molecular targets related to the Wnt/ β -catenin signaling pathway and their potential therapeutic applications.

Keywords: β-catenin, Canonical Wnt, Dishevelled, Frizzled receptor, Glypican-3, Hepatocellular carcinoma, Immunotherapy, Molecular target, Porcupine, Small molecule inhibitor, Tankyrase, Targeted therapy, T-cell factor, Wnt ligand, Wnt signaling.

Atta-ur-Rahman (Ed) All rights reserved-© 2016 Bentham Science Publishers

^{*} Address correspondence to Miran Kim: Liver Research Center, 55 Claverick Street, 4th Floor, Providence, RI 02903, USA; Tel: 401-444-4493; Fax: 401-444-2939; Email: Miran Kim@brown.edu.

INTRODUCTION

Hepatocellular carcinoma (HCC) is the third common cause of tumor-related death (~ 1 million deaths per year) worldwide [1 - 3]. The overall prognosis of patients with liver cancer is poor (mortality: incidence index = 0.95). It is often attributable to a late diagnosis with advanced tumor spread at the time of clinical presentation; thus, most patients have inoperable HCC when they seek medical attention. In this setting, existing therapies have only modest benefit. Among systemic therapies that have been tested with advanced disease, sorafenib, an inhibitor for angiogenesis and Ras-Raf-MAPK signaling, has been shown to significantly effect on patients with HCC due to persistent hepatitis B (HBV) or C (HCV) infection [4]. The growing incidence of HCC and the lack of effective systemic chemotherapy have generated an intense search to unravel the cellular and molecular mechanisms responsible for the pathogenesis of this disease in the hope of developing new innovative approaches [5]. The development of HCC takes a long period of time through a complex multistep process. Accumulation of abnormal genetic and epigenetic alterations as well as deregulation of cellular signaling pathways drive normal hepatocytes or progenitors toward malignant phenotype [6 - 15]. Unlike colorectal cancer in which common genetic changes has been found, HCC displays only few general genetic abnormalities among tumors. Considering various risk factors of HCC, the broad genetic heterogeneity is predictable [16]. Thus, gene expression signature and microRNA profiling have been valuable to categorize patients by distinct subgroups and to define possible driver molecules important in hepatocarcinogenesis [12, 17 - 19]. On the other hand, the extensive genetic heterogeneity of HCC led to focus on determining major cellular signaling pathways that may relate to HCCs with different etiologies and molecular mechanisms. Among these pathways, increasing evidence suggests that impaired Wnt/β-catenin signaling pathway could be critical in hepatic transformation [5, 9].

The Wnt signaling pathway is regulated by multiple components and knowledge of how this pathway is controlled will open new approaches for the development of targeted therapies. In addition to HCC, aberrant activation of this pathway has been shown to be important in a large number of tumors to promote cell growth and appears to control characteristics of the malignant phenotype in various adenocarcinomas of the colorectum [20], breast [21], melanoma [22] and lung [23]. Currently, none of therapeutics targeting the Wnt/ β -catenin signaling has been approved for treatment of HCC yet. It is appealing and great potential to target the Wnt/ β -catenin signaling to treat HCC. Nevertheless, substantial side effects and risks should be considered since the Wnt signaling pathway play a critical role in normal liver homeostasis and regeneration. In this review, we will summarize recent outcomes to intervene the Wnt/ β -catenin signaling pathway and analyze the various therapeutics that target to different stages of the signaling cascade for during HCC development and progression.

THE WNT SIGNALING PATHWAY

Overview of the Wnt Signaling

Wnt signaling regulates diverse developmental and homeostatic functions, including proliferation, differentiation, cell polarity, motility, and migration. The signaling pathways can be classified into two main pathways as canonical (β catenin) and noncanonical pathway. However, the reality is not straightforward to designate Wnt signaling due to crosstalk between two pathways and other factors involved, rather raises complex nonlinear networks. In human, 19 Wnt ligands and 10 Frizzled (FZD) receptors have been identified [24]. Wnt ligands interact with the extracellular domain of FZD receptors through a conserved cystein-rich domain (CRD). This ligand/receptor complex is associated with several regulators, such as the low-density lipoprotein receptor-related protein (LRP) 5/6 co-receptors required for activation of canonical Wnt signaling [25, 26]. In addition, several alternative/cooperative canonical ligands and receptors have been described. In this respect, norrin protein interacts with FZD4 and LRP5 during vascular development [27], and Rspondin-1 can activate the Wnt/β-catenin pathway via Wnt3A in intestinal epithelium [28]. Certain membrane tyrosine kinase receptors can replace FZD receptors; for example, ROR1 and ROR2 interact with the non-canonical Wnt5A and activate the PCP pathway [28, 29]; the Ryk receptor interacts with Wnt1 and Wnt3A of the canonical Wnt [30], and interaction with Wnt5A will activate the non-canonical cascade [31]. Soluble Frizzled-related proteins (SFRPs) can inhibit the Wnt/FZD signal by direct binding with Wnt ligands and by formation of non-functional complexes with

CHAPTER 4

Collaboration of Epithelial Mesenchymal Transition and Cancer Stem Cells: Sinister Routes for Chemoresistant Recurrent Ovarian Cancer

N. Ahmed^{1,2,3,4,*}, K. Abubaker², E. Chan³, G. Kannourakis^{1,5}, J.K. Findlay^{3,4}

¹ Fiona Elsey Cancer Research Institute, Ballarat, Australia

² Department of Surgery, University of Melbourne, Australia

³ Department of Obstetrics and Gynaecology, University of Melbourne, Australia

⁴ Hudson Institute of Medical Research, Melbourne, Australia

⁵ Federation University, Ballarat, Australia

Abstract: Overcoming intrinsic and acquired chemoresistance is the major challenge in treating ovarian cancer patients. Initially nearly 75% of ovarian cancer patients respond favorably to chemotherapy, but subsequently the majority gain acquired resistance resulting in recurrence, cancer dissemination and death. This chapter summarizes recent advances in our understanding of the cellular origin and the molecular mechanisms defining the basis of cancer initiation and malignant transformation with respect to epithelial-mesenchymal transition (EMT) of ovarian cancer cells. We discuss the critical role of EMT frequently encountered in different phases of ovarian cancer progression and its involvement in regulating cancer growth, survival, migration, invasion and drug resistance. Using model ovarian cancer cell lines we highlight the relationship between EMT and the 'cancer stem cell (CSC)-like phenotype' in response to drug treatment, and relate how these processes can impact on chemoresistance and ultimately recurrence. We propose the molecular targeting of distinct 'EMT transformed CSC-like cells' and suggest ways that may improve the efficacy of current chemotherapeutic regimens much needed for the management of this disease.

Keywords: Chemoresistance, Differentiation, EMT, Metastasis, Migration,

* Address correspondence to N. Ahmed: Fiona Elsey Cancer Research Institute, 106-110 Lydiard Street South, Central Park, Ballarat, Vic 3363, Australia; Email: nuzhata@unimelb.edu.au.

Atta-ur-Rahman (Ed) All rights reserved-© 2016 Bentham Science Publishers Ovarian carcinoma, Recurrence, Stem cell markers.

INTRODUCTION

Epithelial ovarian cancer (EOC) is the fifth major cause of cancer mortality in women and constitutes 90% of all ovarian malignancies. In 2012, worldwide there were an estimated 239,000 new cases and over 140,000 deaths from ovarian cancer [1]. Due to lack of specific signs/symptoms and inadequate awareness about the disease on the part of general practitioners as well as the wider community, the disease is not diagnosed until advanced-stages in most patients contributing to a low overall cure rates. The five year survival for stage 1 patients is 90%, but it reduces to 30% in patients with the advanced disease [2]. While surgery is the primary component of initial therapy in advanced-stage patients, almost all patients are treated with chemotherapy to eradicate the residual microscopic and macroscopic peritoneal metastases. Current treatment with paclitaxel and platinum based combination therapy results in a complete remission in 75% patients. Unfortunately, this remission is short lived lasting only for 6-20 months with subsequent recurrence and death as a consequence of metastatic spread [2]. In this chapter we discuss what is known about the initiation of ovarian cancer and the role of epithelial-mesenchymal transition (EMT) in facilitating the progression of this disease. We also shed light on the role of chemotherapyinduced EMT as one of the potential processes that may contribute to the development of chemoresistance and recurrence. In this context, we discuss the relationship between EMT and the acquisition of 'CSC-like phenotypes' in response to chemotherapy. Emphasis has also been made on the development of new therapeutic approaches and how these would impact on EMT and CSC-like phenotypes with the view for better management of this disease.

PATHOLOGY OF OVARIAN CANCER

The term 'ovarian cancer' refers to a diverse group of malignancies that affect the ovaries [3]. Aberrant differentiation is a unique aspect of ovarian cancer biology as tumors acquire the complex differentiation pattern of fallopian tube (serous carcinoma), endometrium (endometroid carcinoma), endocervix (mucinous carcinoma) and vagina (clear cell carcinoma) [4]. Each subtype has identifiable

precursor lesions and multiple early genetic alterations [5]. Hence, mutations in the early progenitor cells may be the contributing factors in the initiation of ovarian carcinoma.

The current classification of ovarian cancer divides the malignancy into two distinct tumor types. Of these, type 1 tumors are low-grade, slow growing tumors, generally confined to the ovary at diagnosis. These tumors develop from well-established precursor lesions commonly termed 'borderline tumors' [3 - 7]. These low-grade ovarian neoplasms have been associated with mutations in KRAS, BRAF, PTEN, and CTNNB1/ β -catenin and include the major histotype, endometrioid, mucinous and low-grade serous carcinomas [3 - 6]. As these genetic alterations occur early in the transformation process, the contribution of these genes defines the tumorigenic process as well as histological differentiation of type 1 tumors.

In contrast, type 2 tumors are high-grade and rapidly progressing tumors which arise from well-defined precursor lesions [6]. Type 2 tumors rarely contain the mutations in KRAS, BRAF, PTEN, and CTNNB1/ β -catenin genes, and vast majority of these neoplasms spread to extra-ovarian sites, specifically, the peritoneum and the fallopian tube early in their development and at a later stage involve the ovary [3 - 6]. Unlike type 1 tumors, type 2 tumors are all high-grade carcinomas, undifferentiated carcinomas or carcinosarcomas [6]. These tumors are composed of large masses of cells with large multinucleated nuclei [6]. They have high mitotic activity and the majority have active DNA damage repair mechanism (DDR) and mutated or ineffective p53 function commonly known as a 'p53 signature' [6, 7]. These tumors may also exhibit gene amplification and over expression of the HER2/neu (10–20%) and AKT2 (10–20%) oncogenes [6]. Gene expression pattern which clusters separately, suggesting that the two groups have a different genetic makeup [8].

The initiating events in EOC are poorly understood. Until recently EOC was thought to arise directly from the surface epithelium of the ovary and its associated inclusion cysts or indirectly from benign ovarian lesions derived from the inclusion cysts [9]. The ovarian surface is composed of a thin layer of surface

CHAPTER 5

Oxaliplatin-mediated Inhibition of Survivin Increases Sensitivity of Head and Neck Squamous Cell Carcinoma Cell Lines to Paclitaxel

P.S. Bisen^{1,*}, Z. Khan², N. Khan³, G.B.K.S. Prasad⁴

¹ School of Studies in Biotechnology, Jiwaji University, Gwalior 474011, India

² Department of Biomedical Sciences and Pathology and Laboratory Medicine, Cedars-Sinai Medical Center, Los Angeles, CA 90048, USA

³ Division of Plant-Microbe Interactions, National Botanical Research Institute, Rana Pratap Marg, Lucknow, 226 001, India

⁴ School of Studies in Biochemistry, Jiwaji University, Gwalior 474011, India

Abstract: Oxaliplatin, is a platinum-based antineoplastic agent used in cancer chemotherapy and paclitaxel is one of several cytoskeletal drugs that target tubulin. Paclitaxel-treated cells have defects in mitotic spindle assembly, chromosome segregation, and cell division. Survivin is one of the key anti-apoptotic proteins which is over-expressed in most of the human cancers; and is associated with their biologically aggressive characteristics and drug resistance. Investigations presented deals with the evaluation of the efficacy of oxaliplatin and paclitaxel combination as a potential strategy in controlling HNSCC cell proliferation and the assessment of correlation between occurrence of apoptosis and changes in expression of survivin (IAP) by employing two HNSCC cell lines (Cal27 and NT8e) and one normal cell line (293) panel with differential level of survivin expression in accordance with chemosensitivity. The combined treatment of cells with paclitaxel and oxaliplatin resulted in a significantly higher cytotoxicity compared to individual single drug treatment. Cytotoxicity was prominent in paclitaxel to oxaliplatin (pacl-oxal) sequence treatment with an approximate two-fold increase in apoptosis compared to oxaliplatin to paclitaxel (oxal-pacl) sequence treatment.

Atta-ur-Rahman (Ed) All rights reserved-© 2016 Bentham Science Publishers

156

^{*} Address correspondence to P. S. Bisen: School of Studies in Biotechnology, Jiwaji University, Gwalior 474011, India; Email: psbisen@gmail.com.

Oxaliplatin-mediated Inhibition

Advances in Cancer Drug Targets, Vol. 3 157

Paclitaxel treatment also significantly increased survivin expression with reduced apoptosis at low concentration. Oxaliplatin, when combined with paclitaxel, decreased the survivin level with increased cell death. Study was further designed to explore the effect(s) of survivin-inhibition by a small interfering RNA (siRNA therapy) method on the apoptosis in HNSCC cells expressing increased sensitivity of the cancer cell lines to paclitaxel whereas over-expression of survivin in the transfected 293-cell line provided resistance. Survivin played a critical role in paclitaxel resistance through the suppression of apoptosis, and a significant induction of apoptosis was observed when oxaliplatin was combined with paclitaxel at least in part by the down-regulation of survivin. In conclusion, the interaction between drugs was synergistic and schedule-dependent.

Keywords: Apoptosis, Chemotherapy, Cytotoxicity, HNSCC, Oxaliplatin, Paclitaxel, Survivin.

INTRODUCTION

In modern oncology, in combination chemotherapy, almost all regimens, the majority of drugs used in cancer chemotherapy are cytostatic in use; combine several chemotherapy drugs, their dosage, the frequency and duration of treatments and other considerations. Chemotherapy regimens are often identified by acronyms, identifying the agents used in the drug combination. Different drugs work through different mechanisms in combination chemotherapy, and that the results of using multiple drugs will be synergistic to some extent because they have different dose-limiting adverse effects. They can be given together at full doses in chemotherapy regimens [1 - 8]. A new approach on combitorial therapeutic relevance of lentiviral mediated survivin knockdown combined the both chemotherapy and ionizing radiation having clinical relevance has been reported which may display a requisite for future combined modality approaches in the treatment of cancer particularly HNSCC (Head and neck squamous cell carcinoma) therapy in cancer treatment with modest effects on cell survival and apoptosis [9]. An approach has been used for identifying the molecular targets of the predicted anti-cancer compounds which were mined from reliable sources like experimental bioassays studies associated with the compound, and from proteincompound interaction databases [7]. Therapeutic compounds from DrugBank, and a list of natural anti-cancer compounds derived from literature mining of

158 Advances in Cancer Drug Targets, Vol. 3

Bisen et al.

published studies, were also used for building partial least squares regression model. The regression model thus built, was used for the estimation of oral cancer specific weights based on the molecular targets [10]. Paclitaxel a microtubulebinding potent cytotoxic agent, treated cells have defects in mitotic spindle assembly, chromosome segregation, and cell division. Paclitaxel has proven clinically efficacious for the treatment of various human cancers including carcinomas of ovarian, breast, head and neck, bladder, lung, and prostate tissues [11 - 15]. There is, however, limited information available on the cellular and molecular mechanisms of the apoptotic effect of paclitaxel on HNSCC cell. Mitosis checkpoint control is important in determining the sensitivity of cancer cell to paclitaxel [16 - 18]. Survivin, a novel anti-apoptotic protein, over-expressed, in most of the human cancers, is reported to be involved in therapy resistance [19 - 24]. We explored the mechanism of HNSCC cell resistance to low concentration of paclitaxel treatment in the present investigations.

Clinically achievable doses of paclitaxel are greatly limited by toxicity to normal tissues, and ranges from sensory neuropathy and gastrointestinal disturbances to severe myelosuppression [25 - 28]. Attempts have been made to overcome this problem by combining paclitaxel with other chemotherapeutic agents [29, 30]. However, paclitaxel-associated neurotoxicity may be exacerbated when used in combination with other neurotoxic agents, such as cisplatin [25, 26]. Therefore, it is important to choose drugs with less toxicity for combination with paclitaxel.

Platinum drugs are one of the important classes of chemotherapy drugs that can induce remissions in various solid tumors [1, 31 - 33]. Oxaliplatin, a third-generation platinum coordination complex, was developed after cisplatin and carboplatin [34, 35]. It lacks such side effects as cisplatin-associated nephrotoxicity and carboplatin induced myelosuppression [34 - 38]. Moreover, clinical studies have demonstrated a chemosensitization effect of oxaliplatin on other drugs, such as 5-FU and folinic acid [39 - 42]. In the present study, the combination of oxaliplatin and paclitaxel was explored as a new strategy for the killing of HNSCC cells and have shown that the addition of oxaliplatin in combination with paclitaxel significantly induced apoptosis, possibly by the down-regulation of survivin.

Melatonin Inhibits the Growth of DMBA-induced Mammary Tumors by Regulating Estrogen Sulfatase Enzyme

Alicia González, Virginia Álvarez-García, Carlos Martínez-Campa, Carolina Alonso-González, Samuel Cos^{*}

Department of Physiology and Pharmacology, School of Medicine, University of Cantabria and Instituto de Investigación Valdecilla (IDIVAL), 39011 Santander, Spain

Abstract: Melatonin has oncostatic effects on different neoplasias, particularly on estrogen-dependent breast cancer, by interacting with estrogen-responsive pathways, thus behaving as an antiestrogenic hormone. In MCF-7 (a human breast adenocarcinoma cell line), melatonin reduces both expression and activity of estrogen sulfatase, thus modulating the local estrogen biosynthesis. In order to investigate the in vivo sulfatase-inhibitory properties of melatonin, this indoleamine was given to ovariectomized rats bearing DMBA-induced mammary tumors also treated with estrone sulfate. In castrated animals, the growth of estrogen-sensitive mammary tumors depends on the local conversion of biologically inactive estrogens to bioactive unconjugated estrogens. Ovariectomy significantly reduced the size and the number of tumors while the administration of estrone sulfate to ovariectomized animals stimulated tumor growth, an effect otherwise abrogated by melatonin. The uterine weight of ovariectomized rats, which depends on the local synthesis of estrogens, was increased by estrone sulfate, being this effect abolished in those animals also treated with melatonin. The growth-stimulatory effects of estrone sulfate on the uterus and tumors depend exclusively on locally synthesized estrogens, since no changes in serum estradiol were observed in estrone sulfate-treated rats. Melatonin counteracted the stimulatory effects of estrone sulfate on sulfatase activity and expression and incubation with melatonin decreased the sulfatase activity of tumors from control animals.

Atta-ur-Rahman (Ed) All rights reserved-© 2016 Bentham Science Publishers

^{*} Address correspondence to Samuel Cos: Department of Physiology and Pharmacology, School of Medicine, University of Cantabria and Instituto de Investigación Valdecilla (IDIVAL), 39011 Santander, Spain; Tel: 34 942 201988; Fax: 34 942 201903; Email: coss@unican.es

182 Advances in Cancer Drug Targets, Vol. 3

González et al.

Animals treated with melatonin had the same survival probability as the castrated animals and significantly higher than the uncastrated. We conclude that melatonin could exert its antitumoral effects on hormone-dependent mammary tumors by downregulating the sulfatase pathway of the tumoral tissue.

Keywords: Breast cancer, DMBA, Melatonin, Pineal, Sulfatase.

INTRODUCTION

Melatonin, the most relevant secretory product of the pineal gland, acts as a regulator of neoplastic cell growth in different kinds of malignant tumors such as prostatic adenocarcinoma, pituitary tumors, leukemia, lung carcinoma, colon carcinoma, neuroblastoma, bladder carcinoma and breast cancer [1 - 3]. The mechanisms that may explain the ability of melatonin to counteract tumor growth were reviewed in Mediavilla et al. [4] and include: antioxidant effects [5], regulation of the estrogen receptor expression and transactivation [6], modulation of the enzymes involved in the local synthesis of estrogens [7 - 9], modulation of cell cycle and induction of apoptosis [10], inhibition of telomerase activity [11], inhibition of metastasis [12], prevention of circadian disruption [13], antiangiogenesis [14 - 16], epigenetic effects [17, 18], stimulation of cell differentiation and activation of the immune system [19]. Perhaps because melatonin was formerly considered as a modulator of the neuroendocrine-reproductive axis, particularly as a controller of seasonal reproduction, the mechanism based on the interplay of melatonin with the estrogen signaling pathway in the context of the hormone-dependent tumors, especially on tumors of the mammary gland, most of which are estrogen-responsive, has been the most thoroughly studied [1 - 3, 20 -22]. Melatonin reduces the estrogen-mediated growth of breast cancer mainly by two different mechanisms: indirect neuroen-docrine mechanisms, through the down-regulation of the neuroendocrine reproductive axis and the consequent reduction of estrogenic hormones responsible for the normal and pathological growth of the mammary gland, and, direct actions of the pineal hormone over tumor cells. In vivo studies on animal models and in vitro studies on human breast cancer derived cell lines support the hypothesis that melatonin oncostatic actions on hormone-dependent mammary tumors are mainly due to its antiestrogenic actions and they are consequence of melatonin interactions on the tumor cells'

Melatonin and Sulfatase in Breast Cancer

estrogen-signaling pathway. On tumor cells, melatonin inhibits the estrogen receptor alpha signaling pathway, thus counteracting the effects of estrogens, therefore behaving as a SERM (selective estrogen receptor modulator). However, melatonin has no effect on estrogen-insensitive breast tumor cell lines [22, 23]. In the last years, research has been focused on the ability of melatonin to regulate the activity of some enzymes (aromatase, sulfatase, 17β -hydroxysteroid dehydrogenase, estrogen sulfotransferase) responsible for the local synthesis of estrogens in cultured human breast cancer cells, thus behaving as a SEEM (selective estrogen enzyme modulator) [24 - 26].

The intratumoral metabolism and synthesis of estrogens may play a very important role in the pathogenesis and growth of hormone dependent breast carcinoma. It is surprising that more than two-thirds of breast cancers occur in postmenopausal women when the ovaries cease to produce estrogens. However, despite the 90% reduction in plasma estradiol levels that occur after the menopause, the concentrations of estrogens in normal breast tissue of pre and postmenopausal women are similar. This can be explained by the extragonadal biosynthesis of estrogens, which occurs in several peripheral tissues, including not only breast, but also adipose tissue, muscle, skin and bone. Estrogens are synthesized in the mammary tissue by transformation either from androgen precursors of mainly adrenal origin or from biologically inactive estrogens [27 -29]. In this situation, plasma levels of estrogens are low, whereas the concentration of estradiol in peripheral tissues such as the mammary gland is high, as a result of the *in situ* biosynthesis [30 - 32]. Yue et al. reported that *in situ* synthesis of estrogen is more important than uptake from plasma to maintain adequate estradiol concentrations in breast tissues after menopause [32]. In fact, in postmenopausal women, it has been demonstrated that intratumoral levels of estradiol are 2- to 3-fold higher than those in areas considered as morphologically normal. A major circulating form of plasma estrogen is estrone sulfate, a biologically inactive precursor of estrogens that might represent a form of reservoir, which can be converted to estrone by the sulfatase, present in both normal and tumoral tissues [33]. Plasma concentrations of estrogen sulfates are 5 to 15 times higher than those of unconjugated estrogens such as estrone, estradiol and estriol, and their half-life in plasma (10-12 hours) is considerably longer than

CHAPTER 7

Role of mTOR Signaling in Tumor Cell Motility, Invasion and Metastasis

Hongyu Zhou¹, Shile Huang^{1,2,*}

¹ Department of Biochemistry and Molecular Biology, Louisiana State University Health Sciences Center, 1501 Kings Highway, Shreveport, LA 71130-3932, USA

² Feist-Weiller Cancer Center, Louisiana State University Health Sciences Center, 1501 Kings Highway, Shreveport, LA 71130-3932, USA

Abstract: Tumor cell migration and invasion play fundamental roles in cancer metastasis. The mammalian target of rapamycin (mTOR), a highly conserved and ubiquitously expressed serine/threonine (Ser/Thr) kinase, is a central regulator of cell growth, proliferation, differentiation and survival. Recent studies have demonstrated that mTOR also plays a critical role in the regulation of tumor cell motility, invasion and cancer metastasis. Current knowledge indicates that mTOR functions as two distinct multiprotein complexes, mTORC1 and mTORC2. mTORC1 phosphorylates p70 S6 kinase (S6K1) and eukaryotic initiation factor 4E (eIF4E) binding protein 1 (4E-BP1), and regulates cell growth, proliferation, survival and motility. mTORC2 phosphorylates Akt, protein kinase C α (PKC α) and the focal adhesion proteins, and controls the activities of the small GTPases (RhoA, Cdc42 and Rac1), and regulates cell survival and the actin cytoskeleton. Here we briefly review current knowledge of mTOR complexes and the role of mTOR signaling in tumor cell migration and invasion. We also discuss recent findings about the mechanism by which rapamycin inhibits cell migration, invasion and cancer metastasis.

Keywords: Akt, Cell motility, Focal adhesion proteins, GTPases, Invasion, Metastasis, mTOR, mTORC1, mTORC2, Rapamycin, S6K1, 4E-BP1.

* Address correspondence to Shile Huang: Department of Biochemistry and Molecular Biology, Louisiana State University Health Sciences Center, 1501 Kings Highway, Shreveport, LA 71130-3932, USA; Tel: (318) 675-7759; Fax: (318) 675-5180; Email: shuan1@lsuhsc.edu

Atta-ur-Rahman (Ed) All rights reserved-© 2016 Bentham Science Publishers

INTRODUCTION

Cancer metastasis, one of the characteristics of malignant tumors, is the primary cause of death in most cancer patients [1]. The metastatic process consists of a series of sequential, interrelated steps including: tumor cells detachment from the primary tumor and invasion of adjacent, healthy tissue, intrusion into the blood and lymphatic vessels, circulation through the bloodstream (circulating tumor cells) to other sites and tissues in the body, extravasation from the vessel of delivery, and metastasis growth in specific distant organs and building a secondary tumor [1 - 3]. Many of these steps are dependent on cell motility and invasion, which allow the cells to change position within the tissues. To spread within the tissues, tumor cells use migration mechanisms similar to that occuring in normal or non-neoplastic cells during physiological processes such as embryonic morphogenesis, inflammatory immune responses, wound healing, and angiogenesis [4]. However, different from the physiological processes of normal cell migration, the tumor cell migration seems to be activated by a variety of promigratory factors without counteracting stop signals, including autocrine motility factors produced by tumor cells, as well as the soluble factors present at the secondary site [5, 6]. Because of this imbalance of signals, cancer cells become continuously migratory and invasive, resulting in tumor expansion across tissue boundaries and the formation of cancer metastasis [6].

Cell migration through tissues results from highly integrated multistep cellular events that are regulated by various signaling molecules, including integrins, Rho family small GTPases, and focal adhesion kinase (FAK) [7 - 9]. In recent years, more proteins and pathways have been identified to be essential for cancer cell motility and invasion, such as phosphatidylinositol 3(kinase (PI3K), protein kinase B/Akt, mammalian target of rapamycin (mTOR), and ribosomal protein S6 kinases (S6K) [10 - 12]. mTOR is known as a key regulator of cell growth, proliferation, differentiation and survival [13, 14]. The mTOR pathway regulates several processes, including protein and lipid synthesis, ribosome biogenesis, autophagy, and metabolism by integrating signals from growth factors, nutrients, oxygen and energy status [14]. Increasing evidence suggests that mTOR pathway also plays an essential role in the regulation of tumor cell motility and invasion, as well as cancer metastasis [11, 15 - 18]. Recent work has identified two

structurally and functionally distinct mTOR-containing multiprotein complexes, mTOR complex 1 (mTORC1) and mTORC2 [14]. The two complexes consist of unique mTOR-interacting proteins which determine their substrate specificity. mTORC1 regulates cell growth, proliferation and survival by promoting anabolic processes, such as protein synthesis, and by limiting catabolic processes [19]. mTORC2 controls the actin cytoskeleton *via* mechanisms by promoting phosphorylation of focal adhesion proteins (focal adhesion kinase, paxillin, and p130^{Cas}) and protein kinase C α (PKC α), and by regulating the activities of small GTPases (RhoA, Rac1 and Cdc42) [20 - 22]. Here, we briefly discuss the insights of mTOR complexes and highlight their roles in the regulation of tumor cell motility and invasion, as well as cancer metastasis. In addition, we summarize recent findings regarding the major mechanisms by which rapamycin inhibit cell migration/invasion and cancer metastasis.

mTOR Strcture and Signaling Complexes

mTOR Structure

mTOR, also known as FRAP (FKBP12-rapamcyin-associated protein). RAFT1 (rapamycin and FKBP12 target), RAPT 1 (rapamycin target 1), or SEP (sirolimus effector protein), is a highly conserved and ubiquitously expressed Ser/Thr kinase [23 - 26]. mTOR and yeast TOR proteins share > 65% identity in carboxyterminal catalytic domains and > 40% identity in overall sequence [27]. mTOR belongs to the PI3K-kinase-related kinase (PIKK) superfamily since the Cterminus of mTOR shares strong homology to the catalytic domain of PI3K [28, 29]. The different members of this family, such as MEC1, TEL1, RAD3, MEI-41, DNA-PK, ATM, ATR, and TRRAP, are associated with diverse cellular functions, such as control of cell growth, cell cycle and DNA damage checkpoints, recombination and maintenance of telomere length [30 - 32]. Structurally, mTOR contains 2549 amino acids and its domain structure is depicted in (Fig. 1). The N-terminus of mTOR possesses up to 20 tandemly repeated HEAT motifs including Huntingtin, elongation factor 3 (EF3), A subunit of protein phosphatase 2A (PP2A), and TOR. The C-terminus consists of the FAT (FRAP, ATM, and TRRAP, all PIKK family members) domain, the FRB (FKBP12-rapamycin binding) domain which is a unique feature of mTOR, a

CHAPTER 8

Structure-Activity Studies on Arylamides and Arysulfonamides Ras Inhibitors

Sonia Colombo, Alessandro Palmioli, Cristina Airoldi, Renata Tisi, Sonia Fantinato, Sandro Olivieri, Luca De Gioia, Enzo Martegani, Francesco Peri^{*}

Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy

Abstract: This paper reports on the synthesis of a panel of small molecules with arylamides and arylsulfonamides groups and their biological activity in inhibiting nucleotide exchange on human Ras. The design of these molecules was guided by structure-activity data previously collected on similar compounds. Aim of this work is the validation of the hypothesis that a phenyl hydroxylamine group linked to a second aromatic moiety generates a pharmacophore capable to interact with Ras and to inhibit its activation. *In vitro* experiments on purified human Ras clearly show that the presence of an aromatic hydroxylamine and a sulfonamide group in the same molecule is necessary to Ras binding and nucleotide exchange inhibition. The inhibitor potency is lower in molecules in which either the hydroxylamine has been replaced by other functional groups or the sulfonamide has been replaced by an amide. In this case both these moieties, the hydroxylamine and sulfonamide are absent, inactive compounds are obtained.

Keywords: Anticancer agents, Computational chemistry, Ras, Structure-activity relationship.

INTRODUCTION

It is well established that K-, H- and N-Ras proteins play a central role in regulating diverse cellular pathways important for cell growth, differentiation and

Atta-ur-Rahman (Ed) All rights reserved-© 2016 Bentham Science Publishers

^{*} Address correspondence to Francesco Peri: Department of Biotechnology and Biosciences, University of Milano-Bicocca, Piazza della Scienza 2, 20126 Milano, Italy; Tel: +39.02.64483453; Fax : +39.02.64483565; Email: francesco.peri@unimib.it.

246 Advances in Cancer Drug Targets, Vol. 3

Colombo et al.

survival [1 - 3]. Deregulation of Ras pathway by activating mutations, overexpression or upstream activation of Ras is common in human tumors [4 - 6]. Of the Ras proteins, K-Ras is the most frequently mutated and is therefore an attractive target for cancer therapy [7, 8]. The complexity of Ras signaling presents many opportunities for therapeutic targeting. A number of different approaches aimed to decrease Ras activity by small molecule inhibitors have been explored in clinical trials [9]. Several of the small molecule therapeutics tested have demonstrated clinical activity, in particular farnesyl transferases inhibitors (FTI), thus supporting the ongoing development of therapies targeting Ras. However, many of the agents currently being evaluated in clinic have multiple targets and their antitumor effects may not be due only to Ras inhibition. To date still no specific and efficient inhibitor of Ras is available for routine clinical use.

In the perspective to develop new molecules directly targeting oncogenic Ras, we have been attracted by small molecules containing the hydroxylamine group and that are able to bind Ras thus interfering with guanine nucleotide exchange, the essential biochemical event leading to Ras activation [10, 11]. We synthesized and tested a first generation of bioactive molecules composed by two aromatic moieties, one of which being a phenylhydroxylamine, connected by a short linker [12]. Some of these compounds showed interesting anticancer properties, unfortunately accompanied by very poor solubility in aqueous buffers and watersolvent mixtures used in biological tests. We therefore decided to project new inhibitors with improved water solubility in which the aromatic moieties able to interact with Ras are linked to polihydroxylated scaffold or polar heterocycles with the aim of improving water solubility [13 - 15]. Some of these compounds were active in inhibiting nucleotide exchange in vitro [16]. Docking experiments pointed out that these compounds bind Ras, and experimental NMR data showed that the two aromatic mojeties on inhibitors are essential for the interaction with Ras [14, 15].

We then developed water-soluble Ras inhibitors by linking natural mono- and disaccharides to phenylhydroxylamine [17]. The interaction between these compounds and Ras was characterized by means of NMR techniques.

We present here a structure-activity characterization of compounds 1-17 as shown

in Fig. (1). This small library of compounds has been generated by progressive structural variations starting from other Ras inhibitors previously developed by us [11] and other groups [9, 10].



Fig. (1). Structural variations on an original common motif: chemical structures of rationally designed Ras ligands 1-17.

The activity in inhibiting *in vitro* the guanine nucleotide exchange and dissociation on purified human Ras and the ability to interfere with the growth of normal mammalian cells and of k-Ras transformed has been evaluated for all compounds. The biological data were complemented with an *in silico* analysis to clarify molecular details of the interaction of these molecules with Ras. In compounds 1-4 a phenylhydroxylamine moiety is linked to either a phenyloxy or a naphtyloxy group through amide or a sulfonamide bonds. In compounds 5-17 the hydroxylamine group has been replaced by other bioisoster groups that mimic some interaction properties of hydroxylamine in terms of hydrogen bond formation (the nitrogen and oxygen atoms of -NHOH group are hydrogen bond donors, while the two hydrogens are acceptors). We therefore replaced hydroxylamine with an amine group in compounds 5 and 8 with an acetamide or

SUBJECT INDEX

4

4E-BP1 207, 211, 213, 218, 219, 221, 222, 226, 227, 229, 232-234

A

Akt 116, 117, 145, 207, 208, 211, 214, 216, 217, 221, 222, 224, 225, 227, 229, 234, 235, 237, 239, 242, 244
Anticancer agents 237, 245
Apoptosis 4, 66, 68, 78, 81, 83, 97, 111, 112, 115, 122, 132, 151, 152, 161, 163, 165, 182, 218, 227, 236, 238, 239, 244

B

Bifunctionality ii, 4

Breast cancer i, iii, 6, 9, 68, 95, 97, 103, 115, 129, 138, 139, 142, 195, 196, 198, 216, 229, 241, 242

С

Canonical Wnt 78, 95, 104

- Cell motility i, iii, 78, 122, 211, 214, 240-242
- Chemoresistance 110, 118, 119, 123, 134, 141, 142, 148, 162, 164
- Chemotherapy 79, 93, 98, 118, 119, 123, 125, 128, 130, 131, 133, 134, 136, 137, 139, 145, 150, 153, 169, 172, 175, 177, 179

Complement resistance 51, 52, 66, 67, 71

Computational chemistry 245, 261

Cytotoxicity ii, 50, 51, 56, 62, 63, 71, 72, 75, 156, 157, 169, 171, 172, 236, 263

D

Differentiation 80, 84, 99, 114, 116, 140, 148, 207, 208, 227, 235, 236, 245

Dishevelled 78, 81, 93, 101, 105, 109, 112, 113 DMBA 181, 182, 185, 196, 197, 200, 203, 205, 206 Dual inhibitors 20, 45

E

Elafin 4, 32, 33, 35, 36, 40-42

Elastokines 4, 7, 8, 32 EMT ii, 118, 119, 135, 137, 139, 141, 142, 147, 152, 218, 222, 241

\mathbf{F}

Flexible docking 4, 33

Focal adhesion proteins 207, 209, 214, 219, 221, 225, 227, 230 Frizzled receptor 78, 107

G

Glypican-3 78, 93, 101, 109, 112 GTPases 214, 219, 222, 223, 227, 242

Η

Hepatocellular carcinoma ii, 78, 79, 116, 141, 155, 218, 221, 229, 237 HNSCC ii, 162, 164, 169, 171-173

Ι

Immunotherapy ii, 50, 51, 66, 71, 72, 78, 92, 111, 154

Invasion i, iii, 4, 6, 43, 78, 84, 118, 123, 124, 214, 239-243

L

Lung cancer i, ii, 13, 15, 19, 20, 26, 42, 45, 92, 101, 103, 111, 149, 169, 174, 226, 236, 238, 243, 244

Atta-ur-Rahman (Ed) All rights reserved-© 2016 Bentham Science Publishers

266 Advances in Cancer Drug Targets, Vol. 3

Μ

Melatonin i, iii, 181-206

Membrane complement regulatory proteins 51 Metastasis iii, 4, 13, 42, 43, 45, 68, 83, 84,

- 96, 100, 118, 125, 129, 130, 132, 138, 139, 152, 154, 155, 174, 182, 214, 221, 223, 236, 241-244
- Migration iii, 14, 39, 80, 84, 107, 115, 118, 124, 130, 133, 135, 153, 218, 240, 242
- Molecular modelling 4, 45
- Molecular target 78, 92, 98
- mTOR i, iii, 244
- mTORC1 207, 226, 227, 230, 231, 234, 237, 238, 240-242 mTORC2 207, 214, 227, 234, 235, 237,
 - 238, 240, 241

N

Neutrophil Elastase ii, 18, 20, 35-49

0

Ovarian carcinoma 125, 146, 150, 151, 153

Oxaliplatin i, ii, 131, 151, 176, 177, 179, 180

P

Paclitaxel ii, 119, 131, 134, 138, 151, 154, 179, 180

Pineal 182, 196, 205, 206 Porcupine 78, 87, 88, 90, 95, 101, 113, 114 Potent angiogenic molecules 7 Potent biheaded Inhibitor 4

R

Rapamycin iii, 174, 229-244

Ras iii, 6, 8, 36, 57, 79, 249-264

Recurrence 38, 67, 100, 108, 118, 119, 123, 133, 134, 139, 154

S

S6K1 207, 216, 226, 227, 229, 233, 234

siRNA 50, 51, 53, 55, 98, 106, 157, 159, 168, 169, 171, 173, 174, 178, 179, 222, 223 Small molecule inhibitor 78, 113, 115, 136

Stem cell markers 119, 126, 128, 136

Structure-activity relationship 21, 245

Sulfatase i, iii, 203-205

Survivin ii, 83, 107, 164-180

Т

T-cell factor 78, 82, 102, 106, 108-110 Tankyrase 78, 86, 87, 90, 96, 98, 115 Targeted therapy 78, 146, 229 Tumor immunology 51

W

Wnt ligand 78, 81, 84, 90, 95 Wnt signaling 90, 94, 95, 98, 100, 102, 103, 105, 123

Atta-ur-Rahman



ATTA-UR-RAHMAN, FRS

Atta-ur-Rahman, Ph.D. in organic chemistry from Cambridge University (1968), has 1020 international publications in several fields of organic chemistry including 727 research publications, 37 international patents, 68 chapters in books and 188 books published largely by major U.S. and European presses. He is the Editor-in-Chief of eight European Chemistry journals. He is Editor of the world's leading encyclopedic series of volumes on natural products "Studies in Natural Product Chemistry" 48 volumes of which have been published under his Editorship by Elsevier during the last two decades.

Prof. Rahman won the UNESCO Science Prize (1999) and was elected as Fellow of the prestigious Royal Society (London) in July 2006. He has been conferred honorary doctorate degrees by many universities including (Sc.D.) by the Cambridge University (UK) (1987), He was elected Honorary Life Fellow of Kings College, Cambridge University, UK, conferred the TWAS (Italy) Prize and the Austrian government has honoured him with its high civil award ("Grosse Goldene Ehrenzeischen am Bande") (2007). He is Foreign Fellow of Chinese and Korean Academy of Sciences, Foreign Fellow of the Chinese Chemical Society and former President of Pakistan Academy of Sciences.