# Frontiers in Natural Product Chemistry

Editor: Atta-ur-Rahman, *FRS* 

**Bentham Books** 

(Volume 6)

Edited by

Atta-ur-Rahman, FRS

Kings College University of Cambridge Cambridge UK

*Volume # 80* 

Editor: Atta-ur-Rahman, HTU

ISBN (Online): 978-981-14-4846-1

ISBN (Print): 978-981-14-4844-7

ISBN (Paperback): 978-981-14-4845-4

©2020, Bentham Books imprint.

Published by Bentham Science Publishers Pte. Ltd. Singapore. All Rights Reserved.

#### BENTHAM SCIENCE PUBLISHERS LTD.

#### End User License Agreement (for non-institutional, personal use)

This is an agreement between you and Bentham Science Publishers Ltd. Please read this License Agreement carefully before using the book/echapter/ejournal (**"Work"**). Your use of the Work constitutes your agreement to the terms and conditions set forth in this License Agreement. If you do not agree to these terms and conditions then you should not use the Work.

Bentham Science Publishers agrees to grant you a non-exclusive, non-transferable limited license to use the Work subject to and in accordance with the following terms and conditions. This License Agreement is for non-library, personal use only. For a library / institutional / multi user license in respect of the Work, please contact: permission@benthamscience.net.

#### **Usage Rules:**

- 1. All rights reserved: The Work is the subject of copyright and Bentham Science Publishers either owns the Work (and the copyright in it) or is licensed to distribute the Work. You shall not copy, reproduce, modify, remove, delete, augment, add to, publish, transmit, sell, resell, create derivative works from, or in any way exploit the Work or make the Work available for others to do any of the same, in any form or by any means, in whole or in part, in each case without the prior written permission of Bentham Science Publishers, unless stated otherwise in this License Agreement.
- 2. You may download a copy of the Work on one occasion to one personal computer (including tablet, laptop, desktop, or other such devices). You may make one back-up copy of the Work to avoid losing it.
- 3. The unauthorised use or distribution of copyrighted or other proprietary content is illegal and could subject you to liability for substantial money damages. You will be liable for any damage resulting from your misuse of the Work or any violation of this License Agreement, including any infringement by you of copyrights or proprietary rights.

#### **Disclaimer:**

Bentham Science Publishers does not guarantee that the information in the Work is error-free, or warrant that it will meet your requirements or that access to the Work will be uninterrupted or error-free. The Work is provided "as is" without warranty of any kind, either express or implied or statutory, including, without limitation, implied warranties of merchantability and fitness for a particular purpose. The entire risk as to the results and performance of the Work is assumed by you. No responsibility is assumed by Bentham Science Publishers, its staff, editors and/or authors for any injury and/or damage to persons or property as a matter of products liability, negligence or otherwise, or from any use or operation of any methods, products instruction, advertisements or ideas contained in the Work.

#### Limitation of Liability:

In no event will Bentham Science Publishers, its staff, editors and/or authors, be liable for any damages, including, without limitation, special, incidental and/or consequential damages and/or damages for lost data and/or profits arising out of (whether directly or indirectly) the use or inability to use the Work. The entire liability of Bentham Science Publishers shall be limited to the amount actually paid by you for the Work.

#### **General:**

- 1. Any dispute or claim arising out of or in connection with this License Agreement or the Work (including non-contractual disputes or claims) will be governed by and construed in accordance with the laws of Singapore. Each party agrees that the courts of the state of Singapore shall have exclusive jurisdiction to settle any dispute or claim arising out of or in connection with this License Agreement or the Work (including non-contractual disputes or claims).
- 2. Your rights under this License Agreement will automatically terminate without notice and without the

need for a court order if at any point you breach any terms of this License Agreement. In no event will any delay or failure by Bentham Science Publishers in enforcing your compliance with this License Agreement constitute a waiver of any of its rights.

3. You acknowledge that you have read this License Agreement, and agree to be bound by its terms and conditions. To the extent that any other terms and conditions presented on any website of Bentham Science Publishers conflict with, or are inconsistent with, the terms and conditions set out in this License Agreement, you acknowledge that the terms and conditions set out in this License Agreement shall prevail.

Bentham Science Publishers Pte. Ltd. 80 Robinson Road #02-00 Singapore 068898 Singapore Email: subscriptions@benthamscience.net



#### CONTENTS

PREFACE	i
LIST OF CONTRIBUTORS	ii
CHAPTER 1 PLANT PROTEIN HYDROLYZATES FROM UNDERUTILIZED	
AGRICULTURAL AND AGROINDUSTRIAL SOURCES: PRODUCTION,	
CHARACTERIZATION AND BIOACTIVE PROPERTIES	
María del Mar Contreras, Minerva Cristina García Vargas, Antonio Lama-Muñoz	
Francisco Espínola, Manuel Moya and Eulogio Castro	
INTRODUCTION	
WAYS OF PRODUCING PLANT PROTEIN HYDROLYZATES FROM	
AGRICULTURAL AND AGROINDUSTRIAL SOURCES	
Sequential Extraction and Hydrolysis (SeEH)	
Extraction by Chemical Methods and Hydrolysis	
Extraction by Enzymatic Methods and Hydrolysis	
Extraction by Physical/Physical-chemical Methods and Hydrolysis	
Simultaneous Extraction and Hydrolysis (SiEH)	
CHARACTERIZATION OF PROTEIN HYDROLYZATES	
Conventional Methods	
Polyacrylamide Gel Electrophoresis	
Capillary Electrophoresis	22
Liquid Chromatography	22
Matrix Assisted Laser Desorption Ionization (MALDI)-MS	
Others	
BIOACTIVE PROPERTIES OF PROTEIN HYDROLYZATES AND ACTIVE PE	
Antioxidant Activity	
Antihypertensive Activity	
Diabetes Targets	
Antimicrobial Activity	
Other Bioactivities	
SAFETY ISSUES	
CONCLUSION	
PATIENT CONSENT	
CONSENT FOR PUBLICATION	
CONFLICT OF INTEREST	
ACKNOWLEDGEMENTS	
REFERENCES	34
CHAPTER 2 NEW DEVELOPMENTS IN THE QUINOLONE CLASS OF ANTIBACT	ERIAL
DRUGS	43
Neslihan Demirbas and Ahmet Demirbas	
INTRODUCTION	43
CLASSIFICATION, SYNTHESIS AND STRUCTURAL REQUIRE-MENTS OF	
QUINOLONE ANTIBACTERIALS	
First Generation	
Second Generation	
Third Generation	
Fourth Generation	
Synthesis of Some Quinolone Drugs	
Rosoxacin	
Norfloxacin, Ciprofloxacin, Fleroxacin	47

THE PROGRESS OF HYBRIDIZATIONS IN QUINOLONES	. 54
Hybridization with Azetidines	
Hybridization with Triazole	. 56
Hybridization with Oxadiazole	. 66
Hybridization with Oxazolidinones and Thiazolidinones	
Hybridization with Thiazole	. 73
Hybridization with Benzimidazole	
Hybridization with Pyrazole	. 75
Oxime Functionalized 4-Quinolones	
Chalcon Functionalized Quinolones	
Hybridization with Metronidazole	
Hybridization with Isatin	
Pyrazine and Isoxacine Hybrids	
Pyrimidine Hybrids	
Hybridization with Flavonoids	
Quinolone-Quinolone Conjugation	
Amidation and Substitution By Homoaromatic Ring at C-3	
Hybridization with β-Lactams	
Hybridization with Macrocycles	
CONCLUDING REMARKS	
CONSENT FOR PUBLICATION	
CONFLICT OF INTEREST	
ACKNOWLEDGEMENTS	
REFERENCES	
ATER PROCESS	108
ATER PROCESS N. Shimizu and T. Ushiyama	
N. Shimizu and T. Ushiyama INTRODUCTION	. 108
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution	. 108 . 108
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure	. 108 . 108 . 110
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles	. 108 . 108 . 110 . 111
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses	. 108 . 108 . 110 . 111 . 112
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION	. 108 . 108 . 110 . 111 . 112 . 121
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION	. 108 . 108 . 110 . 111 . 112 . 121 . 121
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONFLICT OF INTEREST	. 108 . 108 . 110 . 111 . 112 . 121 . 121 . 121
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONFLICT OF INTEREST ACKNOWLEDGEMENTS	. 108 . 108 . 110 . 111 . 112 . 121 . 121 . 121
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONFLICT OF INTEREST	. 108 . 108 . 110 . 111 . 112 . 121 . 121 . 121
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONFLICT OF INTEREST ACKNOWLEDGEMENTS	. 108 . 108 . 110 . 111 . 112 . 121 . 121 . 121
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONFLICT OF INTEREST ACKNOWLEDGEMENTS REFERENCES	. 108 . 108 . 110 . 111 . 112 . 121 . 121 . 121 . 121 . 121
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONFLICT OF INTEREST ACKNOWLEDGEMENTS REFERENCES HAPTER 4 MAJOR METABOLITES OF CERTAIN MARKETED PLANT ALKALOIDS	. 108 . 108 . 110 . 111 . 112 . 121 . 121 . 121 . 121 . 121 . 124
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONFLICT OF INTEREST ACKNOWLEDGEMENTS REFERENCES HAPTER 4 MAJOR METABOLITES OF CERTAIN MARKETED PLANT ALKALOIDS Kuntal Manna, Bikash Debnath and Waikhom Somraj Singh	. 108 . 108 . 110 . 111 . 112 . 121 . 121 . 121 . 121 . 124 . 124
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONFLICT OF INTEREST ACKNOWLEDGEMENTS REFERENCES IAPTER 4 MAJOR METABOLITES OF CERTAIN MARKETED PLANT ALKALOIDS Kuntal Manna, Bikash Debnath and Waikhom Somraj Singh INTRODUCTION	<ul> <li>. 108</li> <li>. 108</li> <li>. 110</li> <li>. 111</li> <li>. 112</li> <li>. 121</li> <li>. 121</li> <li>. 121</li> <li>. 124</li> <li>. 124</li> <li>. 124</li> <li>. 124</li> </ul>
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONSENT FOR PUBLICATION CONFLICT OF INTEREST ACKNOWLEDGEMENTS REFERENCES IAPTER 4 MAJOR METABOLITES OF CERTAIN MARKETED PLANT ALKALOIDS Kuntal Manna, Bikash Debnath and Waikhom Somraj Singh INTRODUCTION Major Metabolites of Certain Important Marketed Alkaloids	<ul> <li>. 108</li> <li>. 108</li> <li>. 110</li> <li>. 111</li> <li>. 112</li> <li>. 121</li> <li>. 121</li> <li>. 121</li> <li>. 124</li> <li>. 124</li> <li>. 124</li> <li>. 125</li> <li>. 125</li> </ul>
<ul> <li>N. Shimizu and T. Ushiyama</li> <li>INTRODUCTION</li> <li>Starch Conformation in Solid State and in Solution</li> <li>Starch Composition and Chemical Structure</li> <li>Preparation of Starch Nanoparticles</li> <li>SEC-MALS, DLS, and Intrinsic Viscosity Analyses</li> <li>CONCLUSION</li> <li>CONSENT FOR PUBLICATION</li> <li>CONFLICT OF INTEREST</li> <li>ACKNOWLEDGEMENTS</li> <li>REFERENCES</li> <li>IAPTER 4 MAJOR METABOLITES OF CERTAIN MARKETED PLANT ALKALOIDS</li> <li>Kuntal Manna, Bikash Debnath and Waikhom Somraj Singh</li> <li>INTRODUCTION</li> <li>Major Metabolites of Certain Important Marketed Alkaloids</li> <li>1. Metabolite Study of Pyridine Group of Alkaloids</li> <li>2. Metabolite Study of Tropane Group of Alkaloids</li> </ul>	<ul> <li>. 108</li> <li>. 108</li> <li>. 110</li> <li>. 111</li> <li>. 112</li> <li>. 121</li> <li>. 121</li> <li>. 121</li> <li>. 124</li> <li>. 124</li> <li>. 124</li> <li>. 125</li> <li>. 126</li> </ul>
<ul> <li>N. Shimizu and T. Ushiyama</li> <li>INTRODUCTION</li> <li>Starch Conformation in Solid State and in Solution</li> <li>Starch Composition and Chemical Structure</li> <li>Preparation of Starch Nanoparticles</li> <li>SEC-MALS, DLS, and Intrinsic Viscosity Analyses</li> <li>CONCLUSION</li> <li>CONSENT FOR PUBLICATION</li> <li>CONFLICT OF INTEREST</li> <li>ACKNOWLEDGEMENTS</li> <li>REFERENCES</li> <li>IAPTER 4 MAJOR METABOLITES OF CERTAIN MARKETED PLANT ALKALOIDS</li> <li>Kuntal Manna, Bikash Debnath and Waikhom Somraj Singh</li> <li>INTRODUCTION</li> <li>Major Metabolites of Certain Important Marketed Alkaloids</li> <li>1. Metabolite Study of Pyridine Group of Alkaloids</li> <li>3. Metabolite Study of Quinoline Group of Alkaloids</li> </ul>	<ul> <li>. 108</li> <li>. 108</li> <li>. 110</li> <li>. 111</li> <li>. 112</li> <li>. 121</li> <li>. 121</li> <li>. 121</li> <li>. 124</li> <li>. 124</li> <li>. 124</li> <li>. 125</li> <li>. 126</li> <li>. 130</li> </ul>
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONSENT FOR PUBLICATION CONFLICT OF INTEREST ACKNOWLEDGEMENTS REFERENCES IAPTER 4 MAJOR METABOLITES OF CERTAIN MARKETED PLANT ALKALOIDS Kuntal Manna, Bikash Debnath and Waikhom Somraj Singh INTRODUCTION Major Metabolites of Certain Important Marketed Alkaloids 1. Metabolite Study of Pyridine Group of Alkaloids 2. Metabolite Study of Tropane Group of Alkaloids 3. Metabolite Study of Quinoline Group of Alkaloids 4. Metabolite study of Isoquinoline Group of Alkaloids	<ul> <li>. 108</li> <li>. 108</li> <li>. 110</li> <li>. 111</li> <li>. 112</li> <li>. 121</li> <li>. 121</li> <li>. 121</li> <li>. 124</li> <li>. 124</li> <li>. 125</li> <li>. 126</li> <li>. 130</li> <li>. 133</li> </ul>
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONFLICT OF INTEREST ACKNOWLEDGEMENTS REFERENCES IAPTER 4 MAJOR METABOLITES OF CERTAIN MARKETED PLANT ALKALOIDS Kuntal Manna, Bikash Debnath and Waikhom Somraj Singh INTRODUCTION Major Metabolites of Certain Important Marketed Alkaloids 1. Metabolite Study of Pyridine Group of Alkaloids 2. Metabolite Study of Tropane Group of Alkaloids 3. Metabolite Study of Isoquinoline Group of Alkaloids 4. Metabolite study of Phenanthrene Group of Alkaloid 5. Metabolite study of Phenanthrene Group of Alkaloid	<ul> <li>. 108</li> <li>. 108</li> <li>. 110</li> <li>. 111</li> <li>. 112</li> <li>. 121</li> <li>. 121</li> <li>. 121</li> <li>. 121</li> <li>. 124</li> <li>. 124</li> <li>. 125</li> <li>. 126</li> <li>. 130</li> <li>. 133</li> </ul>
N. Shimizu and T. Ushiyama INTRODUCTION Starch Conformation in Solid State and in Solution Starch Composition and Chemical Structure Preparation of Starch Nanoparticles SEC-MALS, DLS, and Intrinsic Viscosity Analyses CONCLUSION CONSENT FOR PUBLICATION CONSENT FOR PUBLICATION CONFLICT OF INTEREST ACKNOWLEDGEMENTS REFERENCES IAPTER 4 MAJOR METABOLITES OF CERTAIN MARKETED PLANT ALKALOIDS Kuntal Manna, Bikash Debnath and Waikhom Somraj Singh INTRODUCTION Major Metabolites of Certain Important Marketed Alkaloids 1. Metabolite Study of Pyridine Group of Alkaloids 2. Metabolite Study of Tropane Group of Alkaloids 3. Metabolite Study of Quinoline Group of Alkaloids 4. Metabolite study of Isoquinoline Group of Alkaloids	<ul> <li>. 108</li> <li>. 108</li> <li>. 110</li> <li>. 111</li> <li>. 112</li> <li>. 121</li> <li>. 121</li> <li>. 121</li> <li>. 121</li> <li>. 124</li> <li>. 124</li> <li>. 125</li> <li>. 126</li> <li>. 130</li> <li>. 133</li> <li>. 133</li> </ul>

9. Metabolite Study of Imidazole Group of Alkaloid	
10. Metabolite Study of Terpenoid Group of Alkaloids	
CONCLUSION	
CONSENT FOR PUBLICATION	
CONFLICT OF INTEREST	
ACKNOWLEDGEMENTS	
REFERENCES	145
CHAPTER 5 NATURAL PRODUCTS IN CANCER CHEMOPREVENTION AND	
НЕМОТНЕRАРУ	151
Dev Bukhsh Singh, Manish Kumar Gupta and Rajesh Kumar Pathak	
INTRODUCTION	
MECHANISM OF CANCER	154
ROLE OF NATURAL PRODUCTS IN CANCER	156
Cancer Chemoprevention	162
Cancer Chemotherapy	163
THERAPEUTIC MECHANISM OF ACTION	164
Modulation of Diverse Transcription Factors	164
Targeting Arachidonic Acid Pathway	165
Inhibition of Estrogen Receptor Signaling	166
Targeting Telomerase	166
Promoting the Apoptosis	167
Targeting Receptors (GFRs)	168
Interference with Microtubules	169
Inhibition of Topoisomerase	169
COMBINATION THERAPY OF NUTRACEUTICAL	170
NATURAL PRODUCTS AS A LEAD FOR CANCER THERAPY	171
CONCLUSION	173
CONSENT FOR PUBLICATION	173
CONFLICT OF INTEREST	173
ACKNOWLEDGEMENTS	173
REFERENCES	

# PREFACE

*Frontiers in Natural Product Chemistry* presents recent advances in the chemistry and biochemistry of naturally occurring compounds. It covers a range of topics, including important researches on natural substances. The book is a valuable resource for pharmaceutical scientists and postgraduate students seeking updated and critically important information on bioactive natural products.

The five chapters in this volume are written by eminent authorities in the field. Chapter 1 presents an overview of different ways of production to obtain bioactive peptides from different underutilized plant sources, including from food, brewing and bioethanol industries. Chapter 2 deals with the research on the design of new fluoroquinolones with improved features by molecular hybridization technique. Chapter 3 deals with a pathway for waxy rice starch hydrolysis by a compressed hot water process. Chapter 4 deals with the most important marketed plant alkaloidal drugs and their metabolites. Chapter 5 provides an insight into the molecular basis of preventive and therapeutic effects of natural bioactive substances against cancer diseases.

I hope that the readers will find these reviews valuable and thought-provoking so that they may trigger further research in the quest for new and novel therapies against various diseases. I am grateful for the timely efforts made by the editorial personnel, especially Mr. Mahmood Alam (Director Publications), and Mrs. Salma Sarfaraz (Senior Manager Publications) at Bentham Science Publishers.

Atta-ur-Rahman, FRS Kings College University of Cambridge Cambridge UK

# **List of Contributors**

Ahmet Demirbas	Karadeniz Technical University, Department of Chemistry, Trabzon, Turkey
Antonio Lama-Muñoz	Department of Chemical, Environmental and Materials Engineering, University of Jaén, Campus Las Lagunillas, Jaén, Spain
Bikash Debnath	Natural cum Advance Synthetic Lab, Department of Pharmacy, Tripura University (A Central University), Suryamaninagar, Tripura, India
Dev Bukhsh Singh	Department of Biotechnology, Institute of Biosciences and Biotechnology, Chhatrapati Shahu Ji Maharaj University, Kanpur-208024, India
Eulogio Castro	Department of Chemical, Environmental and Materials Engineering, University of Jaén, Campus Las Lagunillas, Jaén, Spain Center for Advanced Studies in Earth Sciences, Energy and Environment, University of Jaén, Jaén, Spain
Francisco Espínola	Department of Chemical, Environmental and Materials Engineering, University of Jaén, Campus Las Lagunillas, Jaén, Spain Center for Advanced Studies in Earth Sciences, Energy and Environment, University of Jaén, Jaén, Spain
Kuntal Manna	Natural cum Advance Synthetic Lab, Department of Pharmacy, Tripura University (A Central University), Suryamaninagar, Tripura, India
Manish Kumar Gupta	Department of Biotechnology, Faculty of Science, Veer Bahadur Singh Purvanchal University, Jaunpur-222003, Uttar Pradesh, India
Manuel Moya	Department of Chemical, Environmental and Materials Engineering, University of Jaén, Campus Las Lagunillas, Jaén, Spain Center for Advanced Studies in Earth Sciences, Energy and Environment, University of Jaén, Jaén, Spain
Minerva Cristina García Vargas	Department of Chemical, Environmental and Materials Engineering, University of Jaén, Campus Las Lagunillas, Jaén, Spain Department of Industrial Engineering of Tecnológico Nacional de México/Instituto Tecnológico de Zitácuaro, Av. Tecnológico, 186 CP 61534, Zitácuaro, Michoacán, Mexico
Neslihan Demirbas	Karadeniz Technical University, Department of Chemistry, Trabzon, Turkey
N. Shimizu	Research Faculty of Agriculture, Hokkaido University, Hokkaido, 060-8589, Japan Field Science Center for Northern Biosphere, Hokkaido University, Hokkaido, 060-8589, Japan
Rajesh Kumar Pathak	School of Agricultural Biotechnology, Punjab Agricultural University, Ludhiana-141004, Punjab, India
T. Ushiyama	Graduate School of Agriculture, Hokkaido University, Hokkaido, 060-8589, Japan
Waikhom Somraj Singh	Natural cum Advance Synthetic Lab, Department of Pharmacy, Tripura University (A Central University), Suryamaninagar, Tripura, India

### **CHAPTER 1**

# Plant Protein Hydrolyzates from Underutilized Agricultural and Agroindustrial Sources: Production, Characterization and Bioactive Properties

María del Mar Contreras<sup>1,\*</sup>, Minerva Cristina García Vargas<sup>1,2</sup>, Antonio Lama-Muñoz<sup>1</sup>, Francisco Espínola<sup>1,3</sup>, Manuel Moya<sup>1,3</sup> and Eulogio Castro<sup>1,3</sup>

<sup>1</sup> Department of Chemical, Environmental and Materials Engineering, University of Jaén, Campus Las Lagunillas, 23071Jaén, Spain

<sup>2</sup> Department of Industrial Engineering of Tecnológico Nacional de México/Instituto Tecnológico de Zitácuaro, Av. Tecnológico, 186 CP 61534, Zitácuaro, Michoacán, Mexico

<sup>3</sup> Center for Advanced Studies in Earth Sciences, Energy and Environment, University of Jaén, Spain

Abstract: Today, there is a growing interest in the valorization of agricultural and agroindustrial waste/byproducts, including through obtaining bioactive compounds. Besides the use of plant proteins in animal nutrition, obtaining protein hydrolyzates could give an added value, improving digestibility and exerting functional properties by the generation of bioactive peptides. Bioactive peptides encrypted in plant proteins are latent until released and activated by proteolysis. Generally, to obtain bioactive peptides, enzymatic hydrolysis by peptidases is the most common way, with or without previous solubilization and purification steps of the intact protein. This hydrolysis step can be combined with physical and chemical treatments not only to improve the recovery but also to enhance the bioactivity. Therefore, our chapter presents an overview of different ways of production to obtain bioactive peptides from different underutilized plant sources, including from food, brewing and bioethanol industries. In order to characterize bioactive peptides, the application of conventional methods and more sophisticated methods based on mass spectrometry is also described. Moreover, recent literature on the bioactive properties of those plant peptides and current challenges associated with safety issues are discussed.

Keywords: ACE-inhibitor, Antioxidant, Antihypertensive, Antidiabetic, Byproduct, Bioactive peptide, Carbohydrolase, Hydrolysis, Mass spectrometry,

\* **Corresponding author María del Mar Contreras:** Department of Chemical, Environmental and Materials Engineering, University of Jaén, Campus Las Lagunillas, 23071 Jaén, Spain; E-mails: mcgamez@ujaen.es;mmcontreras@ugr.es;mar.contreras.gamez@gmail.com

Atta-ur-Rahman (Ed.) All rights reserved-© 2020 Bentham Science Publishers Microwave assisted extraction, Peptidomics, Peptidase, Protein, Sustainability, Valorization.

#### **INTRODUCTION**

Considering the population factor and the state of natural resources, the need to look for more efficient agroindustry processes is recognized due to demographic growth and the current unsustainable practices. The global population is growing, while our standard of living is increasing; thereby, we have to face environmental challenges. In this sense, it is expected that the world's population increases by 2 billion people in the next 30 years and could reach around 11 billion in the next century. Based on this prognosis, it is not difficult to understand why the United Nation's second priority objective for the present century is to "End hunger, achieve food security and improve nutrition and promote sustainable agriculture" [1]. Moreover, the current agricultural and food practices also threaten the health of people and the planet: i) 70% of worldwide water use is required by agriculture; ii) it generates huge levels of pollution and waste; iii) risks associated with poor diets are one of the leading causes of death; iv) a double burden of malnutrition exists since millions of people are either eating not enough or eating the wrong types of food. In 2017, this led to one in eight adults (*i.e.* more than 672 million people) in the world to be obese [2] and forecasts suggest high levels of obesity on the future population [3]. In particular, increased demand for animalbased protein is expected to have a negative environmental impact, generating greenhouse gas emissions, requiring more water and more land [4]. Thereby, plant proteins could be an alternative but sustainable practices are required.

Therefore, against this background, the goal is how to meet the growing global demand for food, including protein and healthy foods, to improve income and employment in rural areas and, at the same time, reduce the environmental impact. This puts pressure on the world's resources to provide not only more but also different types of food, including more sustainable production of existing sources of protein as well as alternative sources for human consumption [4]. This requires us to move from an oil based economy towards a more sustainable circular bioeconomy model, producing more food and bio-based products from renewable resources, including agricultural and agroindustrial byproducts [5]. In this line, the biorefinery concept has emerged as a sustainable processing of biomass into a portfolio of marketable food and feed ingredients, bio-based products (chemicals, materials, proteins, bioactive compounds, *etc.*) and energy (fuels, power, heat) [6 - 8].

When thinking about these resources, plant compounds are usually put forward as their most probable source [9]. This includes macro (cellulose, hemicelluloses,

#### Plant Protein Hydrolyzates

#### Frontiers in Natural Product Chemistry, Vol. 6 3

pectins, starch, lignin, proteins, minerals, etc.) and microcomponents (e.g. phytochemicals). In particular, plant byproducts are underutilized sources of proteins and, most of the time are addressed to animal nutrition, but the ruminal degradability of proteins is not high. Nonetheless, proteins can be beneficial not only in terms of nutrition but also from a functional point of view through the generation of bioactive peptides. This means that the breakdown of peptide bonds by enzymatic hydrolysis increases the solubility, digestibility, and functional properties of the precursor proteins and byproduct [6]. Bioactive peptides are known for their high tissue affinity, specificity and efficiency in promoting health [10]. Therefore, apart from the use of plant proteins in animal nutrition, obtaining protein hydrolyzates could give an added value in a biorefinery context, with improved digestibility and exerting functional properties through the generation of bioactive peptides. This could also lead to the formulation of functional ingredients that are in line with the increased consumer awareness towards functional foods, nutraceuticals and personalized diets; the driving force of the functional food and nutraceutical market [10]. Moreover, there is a growing interest in the food industry and among consumers in reducing the use of synthetic additives in food preservation and opting instead for natural ones [11]. All this together connects with the concept of bioeconomy since it can promote a new way to diversify plant byproducts.

Generally, hydrolysis by peptidases, with or without a previous protein extraction step, is the most common way to obtain bioactive peptides with a wide range of biological properties. e.g. antidiabetic, antihypertensive, antimicrobial. antioxidant, and anticancer properties [12 - 15], but also autolysis and application of microbial suspensions (whole cells) have been applied [13, 16]. Enzymatic hydrolysis can be combined with physical treatments and alkaline extraction not only to improve the recovery but also to enhance the bioactivity [6, 17]. In this context, this book chapter presents an overview of the different ways of production to obtain bioactive protein hydrolyzates from different underutilized plant sources. These sources include byproducts from the cereals industry (wheat germ protein, broken rice by-product), oil industry (olive, and rapeseed/canola byproducts), fruit and vegetable industries (e.g. fruits seeds, potato byproducts, cauliflower leaves), and brewing industry (brewer's spent grain). Some techniques applied to characterize the hydrolyzates and the peptides are also covered. Moreover, the biological properties of the hydrolyzates have been revised, and the sequence of some bioactive peptides is shown. Finally, some safety issues are also discussed.

43

**CHAPTER 2** 

# New Developments in the Quinolone Class of Antibacterial Drugs

#### Neslihan Demirbas<sup>\*</sup> and Ahmet Demirbas

Karadeniz Technical University, Department of Chemistry, 61080 Trabzon, Turkey

Abstract: The increasing drug resistance and the insufficiency of the newly developing antibiotics constitute a serious and growing health threat in the world. Especially Gram (-) bacteria acquire genetic material encoding antibiotic resistance by multiple mechanisms. Development of novel antibacterial agents with little tendency to bacterial resistance is, therefore, an important and challenging topic in the medicinal chemistry, and synthetic organic chemistry is an indispensable part of the design and synthesis of efficient antibacterial drug candidates. Among the broad-spectrum antibiotics, fluoroquinolones constitute the most attractive drugs in the anti-infective chemotherapy field. These antibiotics target the bacterial type II topoisomerase enzymes (DNA gyrase and topoisomerase IV) which are essential enzymes involved in bacterial cell growth and division. Since their advent, they were widely applied to treat infections. Unfortunately, most of them suffered from the resistance problem by mutations in the bacterial targets due to their wide use. Recently, the synthetic organic and medicinal chemists focused their research on the design of new fluoroquinolones with improved features by molecular hybridization technique. One of the most promising approaches aiming to combat resistant pathogens is the design and synthesis of new hybrid molecules in which different pharmacophore groups with different modes of action are joined together using a flexible linker. This strategy supplies a way to improve traditional drug combination therapies simplifying optimization of the pharmacokinetics/pharmacodynamic (PK/PD) profile, efficacy at both targets is usually synergistic.

**Keywords:** Aminoglycoside, Drug resistance, Flavonoid,  $\beta$ -Lactam, Macrocyclic, Molecular hybridization, Oxazolidinone, Pyrazole, Pyrazine, Pyrimidine, Quinolone, Triazole.

#### INTRODUCTION

In recent years, the growing incidence of virulent bacterial resistance towards the present antibacterial agents has become the most serious clinical and socio-

Atta-ur-Rahman (Ed.) All rights reserved-© 2020 Bentham Science Publishers

<sup>\*</sup> Corresponding author Neslihan Demirbas: Karadeniz Technical University, Department of Chemistry, 61080 Trabzon, Turkey; Tel/Fax: +90 462 3774252; E-mail: neslihan@ktu.edu.tr

economic problem worldwide [1 - 3]. Although, The World Health Organization, has described the antibiotics as "miracle weapons giving an opportunity to combat with infectious diseases", a large majority of clinically effective drugs actively used to treat bacterial infections have become less effective due to the increasing antimicrobial resistance [4 - 9]. Moreover, the treatment of infectious diseases is more difficult in immunodeficient patients, such as those infected with tuberculosis, HIV *etc* [9]. Multidrug resistant Gram (+) pathogens, such as methicillin-resistant Staphylococcus aureus (MRSA) and *Staphylococcus epidermis* (MRSE), vancomycin-resistant *Enterococci* (VRE), cephalosporin resistant *Streptococcus pneumoniae* are leading significant morbidity and mortality of the infected patients [10 - 12]. Another pathogen, penicillin resistant *S. pneumoniae* has been reported to cause approximately 3 million deaths each year worldwide because of pneumonia, meningitis and sepsis, which are responsible for serious upper airway infections, such as sinusitis and otitis media [13 - 16].

Microorganisms develop resistance to drugs *via* various mechanisms, such as overexpression of drug efflux transporters, like multidrug and toxic compound extrusion (MATE) transporters [17], changes in the target sites of antibiotics [18], optimization of the enzyme (such as  $\beta$ -lactamase) activity resulting in inactivation of antibiotics [19], spontaneous chromosomal mutations [20], and horizontal transfer of genetic elements [21]. Inhibition of the activity of drug efflux transporters appears to be an encouraging strategy for renovating the activity of a drug that is the substrate of these efflux pumps [22].

Keeping all this in mind, it is clearly seen that the development of wholly novel drug discovery methodologies and the optimization of available antibacterial agents have become a crucial and challenging task for the effective treatment of bacterial infections. However, the development of completely new antibacterials suitable for therapeutic applications has not been as successful as expected, and despite a tenfold increase in spending for Research-Development studies in the pharmaceutical industry, the number of leader molecules has remained nearly stable.

To improve the therapeutic profile of the existing drugs by several manipulations in their structures or to design their novel analogs has become one of the most promising strategies for the development of new antibacterial drugs. This strategy has been widely admitted since it does not entail to discover novel scaffolds or validation of new biological targets, which has been accepted as an extremely difficult and time-consuming procedure [27].

In recent years, in order to overcome the "drug resistance nightmare", the concept

of "molecular hybridization" based on the combination of structural features of two or more drug fragments having different modes of action has emerged as an attractive strategy. These new hybrid compounds with improved affinity and efficacy have been proved to be capable of inhibiting two or more conventional targets simultaneously, and this multiple target strategy has led to discover a number of bioactive hybrid molecules [28 - 31].

In recent drug development programs, 4-quinolone-3-carboxylic acid scaffold has been used as one of the most frequently encountered privilege frameworks having potent and broad spectrum activity [32]. Since the introduction of nalidixic acid (the first generation, the prototype 4-quinolone antibiotics) for the treatment of urinary tract infections in humans in 1962, the class of quinolone antibacterials has played an important role saving countless millions of lives in the chemotherapy of bacterial infections [33 - 35]. Their preferable properties including well tolerability with excellent safety profile, favorable pharmacokinetic characteristics, broad antibacterial spectrum and good treatment effectiveness have made quinolones an important class of synthetic antibacterial agents [36, 37]. This class of antibacterials displays direct inhibition activity on the DNA synthesis by binding to the enzyme DNA complex, they stabilize DNA strand breaks created by DNA gyrase and topoisomerase IV [38]. Gyrase is responsible for introducing negative supercoils in DNA and relieving torsional stress expected to accumulate ahead of transcription and replication complexes. Topoisomerase IV provides a potent decatenating activity. Both gyrase and topoisomerase IV are essential enzymes and therefore the compounds that block bacterial growth by inhibiting them are accepted as potential chemotherapeutics [39].

#### CLASSIFICATION, SYNTHESIS AND STRUCTURAL REQUIRE-MENTS OF QUINOLONE ANTIBACTERIALS

Until today, four generations of quinolone class antibacterial drugs have been developed.

#### **First Generation**

The first generation includes: nalidixic acid, oxolinic acid, pipemidic acid, cinoxacin and rosoxacin which have shown only weak to moderate activity against Gram (-) bacteria, and therefore is rarely used today.

# **CHAPTER 3**

# Structure of Fine Starch Prepared *Via* a Compressed Hot Water Process

N. Shimizu<sup>1,2,\*</sup> and T. Ushiyama<sup>3</sup>

<sup>1</sup> Research Faculty of Agriculture, Hokkaido University, Hokkaido, 060-8589, Japan

<sup>2</sup> Field Science Center for Northern Biosphere, Hokkaido University, 060-0811, Japan

<sup>3</sup> Graduate School of Agriculture, Hokkaido University, Hokkaido, 060-8589, Japan

Abstract: In a "top-down" process, starch nanoparticles can be produced by structural and size refinements through the breakdown of large particles. In this study, the structure of fine starches prepared via a compressed hot water process at different temperatures  $(160 - 180^{\circ}C)$  was analysed using dynamic light scattering and sizeexclusion chromatography with multi-angle light scattering (MALS) and differential refractive index detection. Changes in the molecular weight, polydispersity, hydrodynamic radius, and radius of gyration were assessed. The intrinsic viscosity of the fine starch solution was derived from the Flory-Fox and Ptitsyn-Eizner equation. The weight-average molecular weight decreased to  $7.29 \times 10^{-5}$  g/mol while the average hydrodynamic radius and weight-average radius of gyration decreased by 34.9 nm and 14.6 nm respectively, in fine starch prepared at 180 °C. In fine starches prepared at 160 °C, 165 °C, and 170 °C, tails in the multi-angle light scattering peaks, upswings in the conformation plots, and upturns in the plots of gyration radii and elution volumes were all the result of branching structures. In fine starches prepared at 175 °C and 180 °C, amylopectin branching was diminished and symmetrical scattering peaks were detected in the MALS analysis. We propose a pathway for waxy rice starch hydrolysis by a compressed hot water process.

**Keywords:** Fine starch, Intrinsic viscosity, Hydrodynamic radius, Molecular weight, Radius of gyration.

#### INTRODUCTION

#### Starch Conformation in Solid State and in Solution

Starch is a renewable and biodegradable biopolymer that is stored in many plants as a source of energy for photosynthesis. It is the second most abundant biomass in nature, and is typically isolated from plants in the form of microscale granules.

\* Corresponding author Shimizu N: Research Faculty of Agriculture and Field Science Center for Northern Biosphere, Hokkaido University, Japan; Tel/Fax: +81-11-706-3848; E-mail: shimizu@bpe.agr.hokudai.ac.jp

Atta-ur-Rahman (Ed.) All rights reserved-© 2020 Bentham Science Publishers

#### Structure of Fine Starch Prepared

Fine starches with average particle sizes ranging from the micrometre to nanometre scale have been developed as functional materials with applications in foods, cosmetics, medicines, and various composites. The major characteristics of fine starches are rapid dissolution and enhanced bioavailability after consumption.

Recent studies have reported that nano-scale starch particles can be readily prepared from starch granules, which have unique physical properties [1]. Starch granules consist of numerous nano-size semi-crystalline blocklets [2]. Physical treatments may disintegrate the starch granules, thus releasing the nano-blocklets. The preparation of starch nanoparticles may be classified into "top-down" and "bottom-up" processes. In the "top-down" process, nanoparticles are produced from structural and size refinement through the breakdown of large particles [3, 4]. In the "bottom-up" process, starch nanoparticles self-assemble into starch particles. Starch nanoparticles are important vehicles for nano- and microencapsulation in the food industry [5 - 7]. The physicochemical properties of polymers such as molecular weight, polydispersity, radius of gyration, hydrodynamic radius, and the molecular structure in solution are derived before and after modification processes [8, 9]. The intrinsic viscosity of the fine starch solution can be derived using the Flory-Fox and Ptitsyn-Einzer equation. It is important to evaluate not only raw starch, but also modified starches, for industrial use [3, 4].

To use starch effectively, techniques to prepare nanoscale starches have been developed and assessed. Nano-scale waxy rice starch particles can be prepared *via* hydrolysis using a compressed hot water process. Starch nanoparticles can also be prepared by acid hydrolysis, enzymatic treatments, and physical treatments such as high-pressure homogenization, ultrasonication, reactive extrusion, and gamma irradiation [1]. The compressed hot water process is one of the most useful reactions. The smallest average hydrodynamic radius of 75.2 nm was obtained by using a 180°C compressed hot water treatment, with a starch concentration of 0.1% (w/w), and an initial pressure of 3.0 MPa. The product of this process was evaluated by zeta potential and by using a submicron particle size analyzer [10]. A fine waxy rice starch solution prepared at 160°C with a compressed hot water treatment was spray-dried as a wall material for micro encapsulation [6]. It is important to understand starch macromolecular structures to optimize the practical uses of various products in industry. However, the changes in the hydrodynamic particle size induced by the compressed hot water process are unknown.

In this chapter, we describe the structure of fine starches prepared *via* a compressed hot water process at different temperatures  $(160^{\circ}C - 180^{\circ}C)$ . The fine starches were analysed using dynamic light scattering (DLS) and size-exclusion chromatography (SEC) with multi-angle light scattering (MALS) and differential

refractive index (DRI) detection.

The intrinsic viscosity was calculated using the Flory-Fox and Ptitsyn-Eizner equation and correlations with particle conformation were derived. In SEC-MALS measurements, the branching amylopectin polymers affected the column separation and the MALS signal. Therefore, changes in the branching structures in waxy rice starch by the compressed hot water process could be deduced from the SEC-MALS data.

#### **Starch Composition and Chemical Structure**

Starch is commonly extracted from corn, wheat, and tapioca in native and modified forms for applications in the food, paper, and pharmaceutical industries [11 - 14]. After extraction from plants, starch occurs as flour-like white particles that are insoluble in cold water. For example, rice starch granules are semicrystalline particles ranging from 3 to 8  $\mu$ m. The smallest known starch particles are those in cereal grains. There is some variation in starch granule size among different rice genotypes. Rice starch granules have a smooth surface, but angular and polygonal shapes. The granules are loosely packed in clusters, and some particles have holes and cracks (Fig. 1).

The internal architecture of native starch granules is characterized by "growth rings" that represent concentric semi-crystalline shells (thickness 120 - 400 nm) separated by amorphous regions. There is evidence that the crystalline shells consist of regular alternating amorphous and crystalline lamellae repeating at 9 - 10 nm intervals. In this structural organization, parallel double helices of amylopectin side chains assemble into radially oriented clusters (Fig. 2). Little is known about the structure, organization, and arrangement of the lamellae.

Starch consists of amylase  $\alpha(1-4)$ -linked glucose units, amylopectin  $\alpha(1-4)$ -linked glucose units, and branched  $\alpha(1-6)$ -linkages. The molecular weights ( $M_w$ ) of amylose and amylopectin in starch vary among different plants; in normal corn they are  $1.4 \times 10^6$  and  $39 \times 10^6$ g/mol, respectively [15]; that of amylose from rice is  $5.1-6.9 \times 10^5$  g/mol [16]; that of amylose from waxy barley starch is  $1.06 \times 10^8$  g/mol [17]; and that of amylose in Amioca (waxy corn starch) ranges from 107-10<sup>9</sup> g/mol [18].

Starch molecules have a semi-crystalline structure that significantly affects their physical and chemical performance. Analyses of the fine waxy starch after the compressed hot water process revealed a peak derived from its crystalline structure. This peak was much reduced after the hydrothermal and spray-drying processes, indicating that the crystalline structure in the starch molecules had broken down into the amorphous form.

# Major Metabolites of Certain Marketed Plant Alkaloids

#### Kuntal Manna<sup>\*</sup>, Bikash Debnath and Waikhom Somraj Singh

Natural cum Advance Synthetic Lab, Department of Pharmacy, Tripura University (A Central University), Suryamaninagar, Tripura, India

**Abstract:** The archeological and historical record shows that people across Asia, Europe, and Africa used alkaloidal drugs as early as 2000 BCE. Alkaloids are heterocyclic rings consisting of at least one nitrogen atom. They are the waste products of plant metabolites and serve a wide variety of biological activities to human beings. Nicotine, cytosine, atropine, scopolamine, cocaine, catuabine, quinine, quinidine, dihydroquinine, papaverine, ephedrine, reserpine, ergotamine, caffeine, *etc.* are the most important marketed plant alkaloidal drugs and their metabolites are described in this chapter. Metabolism plays a central role in regulating the toxicity of a variety of phytochemicals. Hepatic microsomal enzymes such as monooxygenase and putative NADPH-FMN-reductase, carboxyl esterase, CYP2B6, CYP3A4, and CYP2D6 are mostly involved in the metabolism of alkaloids. This chapter will be important for future researchers.

**Keywords:** Cytochrome P650, Heterocyclic Ring, Hepatic Microsomal Enzymes, Marketed Alkaloids, Major Metabolites , Metabolic Pathway, Pharmacological Activities, Secondary Metabolites.

#### **INTRODUCTION**

Natural medicines provide a major source of pharmaceuticals, which we use today directly from nature or in marketed form. For the assistance of plants to survive and reproduce, they synthesize many secondary metabolites [1]. These secondary metabolites reveal biodynamic activity beneficial to both human and animal health. Alkaloids, phenols, steroids, glycosides, tannins, terpenoids, and phytoalexins are the secondary metabolites produced by the plants [2]. Among these secondary metabolites, alkaloids are considered the important ones. They are relatively modest molecules existing in plants at <10 g/kg [3]. Due to being toxic in nature, plants use alkaloids to protect themselves against harmful organi-

<sup>\*</sup> **Corresponding author Kuntal Manna:**Department of Pharmacy, Tripura University (A Central University), Suryamaninagar, Tripura, India-799022; Tel: +91381-2379404; Fax: +91381-2374803; E-mail: k manna2002@yahoo.com

#### Major Metabolites

sms. Alkaloids are constituted in the plant kingdom and mainly found in the higher plants, such as those belonging to papaveraceae, menispermaceae, ranunculaceae, leguminosae, and loganiaceae [4, 5]. Groups of nitrogen-containing compounds that may consist of one or more nitrogen atoms (within the heterocyclic ring) are called alkaloids. However, the term 'alkaloid' (alkali-like) is rather interesting as there is no definite borderline between alkaloids and naturally existing complex amines. Typical alkaloids are basic in nature and primarily acquired from plant sources [6, 7]. Alkaloidal drug metabolism can have significance due to its therapeutic effect or its toxicity. It mainly takes place in the liver and the Cytochrome P450 enzymes are involved in a vital role in metabolism [8]. This chapter mainly focuses on biological sources along with major metabolites in some important marketed alkaloid drugs.

#### Major Metabolites of Certain Important Marketed Alkaloids

#### 1. Metabolite Study of Pyridine Group of Alkaloids

**Nicotine 1** (Fig. (1) is a naturally occurring alkaloid found in many plants. Dried leaves of Nicotiana tabacum belonging to the family Solanaceae constitute the active source of nicotine [9]. Nicotine and its metabolites may be dangerous to the body. Hepatic enzyme Cytochrome P450 2A6 (CYP2A6), UDPglucuronosyltransferase (UGT), and flavin-containing monooxygenase (FMO) play an active role in nicotine metabolism. Electrospray ionization and highperformance liquid chromatography/tandem-mass spectrometry (LC-MS/MS) methods are used for the determination of nicotine metabolites Fig. (1). Quantitatively, the most important metabolites of nicotine in mammals are the lactam derivative and cotinine. In humans, about 70-80% of nicotine is converted into cotinine. Nicotine N'-oxide is another primary metabolite of nicotine, although only about 4–7% of nicotine consumed by active smokers is metabolized via this route. The kidney is the vital organ for the excision of nicotine [10, 11]. **Cytisine 26** Fig. (2) is a selective nicotinic cholinergic agonist alkaloid obtained from the seed of *Laburnum anagyroides* belonging to the family Fabaceae [12]. Astroug et al. (2010) described the pharmacokinetics of cytisine. They gave oral and intravenous administration to examine the cytisine pharmacokinetics profile in male and female New Zealand rabbits. After the administration of cytisine, the rabbit serum of both the sexes were collected in a specific time interval to measure the pharmacokinetic parameters using high-performance liquid chromatography (HPLC) method with ultraviolet (UV) detection. The pharmacokinetic analysis suggested a rapid but incomplete absorption of cytisine after oral administration and did not clarify any metabolites of cytisine [13]. Later Jeong et al. (2015) developed a liquid-chromatography mass spectrometry (LCMS) method for the pharmacokinetic study of cytisine in human plasma and

urine. No metabolites were detected in plasma or urine collected in their study [14]. Further research is required for a better understanding.

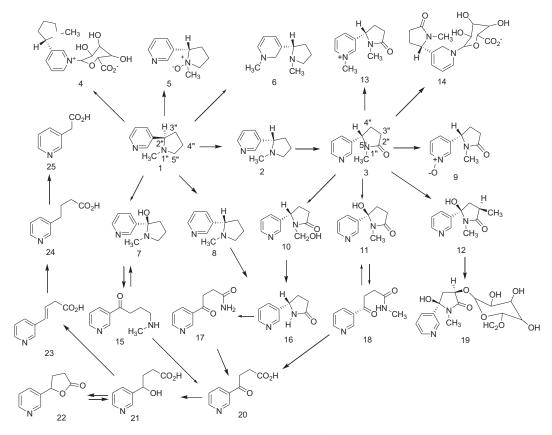


Fig. (1). Metabolism pathway of nicotine (1), Nicotine  $\Delta^{1/(5')}$  iminium ion (2), cotinine (3), Nicotine glucoronide (4), Nicotine N'- oxide (5), Nicotine isomethonium ion (6), 2- hydroxy nicotine (7), Nornicotine (8), Cotinine N- oxide (9), N'- hydroxymethyl norcotine (10), 5- hydroxycotinine (11), *Trans-3'*-hydroxycotinine (12), Cotinine methonium ion (13), Cotinine glucorinide (14), 4- (methylamino)-1- (3-pyridyl)-1-butanone (15), Norcotine (16), 4-oxo-4- (3-pyridyl)-butanamide (17), 4-oxo-4(3-pyridyl) N-methylbutanamide (18), *Trans-3'*-hydroxycotinien glucoronide (19), 4-oxa-4(3-pyridyl)-butanoic acid (20), 4-hydroxy-4-(3-pyridyl) butanoic acid (21), 5-(3-pyridyl)-tetrahydrofuran-2-one (22), 4-(3-pyridyl)-3-butanoic acid (23), 4-(3-Pyridyl)-butanoic acid (24), 3-Pyridylacetic acid (25).

#### 2. Metabolite Study of Tropane Group of Alkaloids

Atropine 27 Fig. (3) is obtained from the plant *Atropa belladonna*, a perennial herb belonging to the family Solanaceae. For intoxication in nature, this drug typically provides anticholinergic effects to the body [15]. Atropine is metabolized in the liver, and 30-50% of its unchanged are excreted along with urine [16]. Chen *et al.* (2006) outlined a metabolic pathway of atropine from rat urine, based on the LC-MS/MS technique after the administration of atropine

# Natural Products in Cancer Chemoprevention and Chemotherapy

Dev Bukhsh Singh<sup>1,\*</sup>, Manish Kumar Gupta<sup>2</sup> and Rajesh Kumar Pathak<sup>3</sup>

<sup>1</sup> Department of Biotechnology, Institute of Biosciences and Biotechnology, Chhatrapati Shahu Ji Maharaj University, Kanpur-208024, India

<sup>2</sup> Department of Biotechnology, Faculty of Science, Veer Bahadur Singh Purvanchal University, Jaunpur-222003, Uttar Pradesh, India

<sup>3</sup> School of Agricultural Biotechnology, Punjab Agricultural University, Ludhiana-141004, Punjab, India

Abstract: Cancer is a very fatal, challenging and complex disease. A large number of people across the globe are suffering from various types of cancer. The bioactive substances isolated from different parts of several herbs and spices have shown their valuable preventive and therapeutic role against different forms of cancer. The recent technological innovations have made it possible to explore the molecular targets of these bioactive substances and also enabled us to know the mechanism of action related to disease modulation. Our traditional knowledge related to chemopreventive and chemotherapeutic role of herbs is now being validated and explored by the use of modern biological techniques. However, mechanism of action that governs anticancer effect has not been well elucidated for many herbs and natural products. Herbal extracts are the mixture of different active substances, therefore screening and pharmacological response of each individual compound should be validated separately to gain some insight about mechanism of anticancer property. Polyphenols play significant role in initiation, promotion and progression of cancers by modulating the enzymes and signal of diverse pathways related to cellular proliferations, differentiation, angiogenesis, apoptosis and metastasis. These natural bioactives also serve as lead compounds for drug designing and their biological activity can be further optimize by some desired chemical modification. Optimisation of the therapeutic inhibitors can be enhanced using systems biology modelling of the molecular pathways related to the disease. The aim of this chapter is to provide the insight into the molecular basis of preventive and therapeutic effect of natural bioactive substance against cancer diseases.

**Keywords:** Apoptosis, Cancer, Chemotherapy, Combination therapy, Metastasis, Natural products, Polyphenols Cell division, Telomerase, Topoisomerase, Transcription factor.

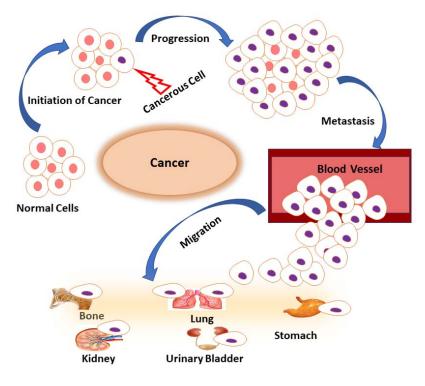
<sup>\*</sup> Corresponding author Dev Bukhsh Singh: Department of Biotechnology, Institute of Biosciences and Biotechnology, Chhatrapati Shahu Ji Maharaj University, Kanpur-208024, India; E-mail: answer.dev@gmail.com

#### INTRODUCTION

Cancer is a very challenging disease for researchers to find an effective approach for the detection and treatment. Cancer is the result of genetic changes that transform the normally dividing cells into malignant cells. It is an uncontrolled division of cells, where cells divides rapidly without reaching to stage of maturity. There are different types of cancer based on the parts of body and mechanism of disease. Cancer is the leading cause of premature death and physical disability world-wide and adversely affects the social and economic status of a country [1]. General causes of cancer are toxic chemicals, radiations, pathogens and genetic factors. Sign and symptoms of cancers are specific to the parts of body where it develops. Many cancers are identified by the name of body parts where abnormal growth of cell or tissues occurs such as breast cancer, colon cancer, lung cancer, liver cancer, prostate cancer, thyroid cancer and pancreatic cancer. Some of the general signs and symptoms are weight loss, pain, bleeding, fever, persistent cough and unusual tissue masses.

Despite of having advancements in screening, detection and therapeutic approaches for the treatment of cancer, the burden of cancer is expected to increase in future. Major cause of cancer death is the lack of detection in early stage. Treatment strategies for cancer depends on the type and stage of cancer. Natural products have proven their effective role in the prevention of different types of cancer. Natural products have also shown their potential role in treatment of cancer [2]. Modern techniques of biology have played very important role in elucidating the mechanism of action behind the preventive and therapeutic role of herbal products. With the discovery of mechanism that are involved in progression of a cancer, designing of a therapeutic molecule targeting the enzyme can be initiated. Surgery, chemotherapy and radiation therapy are the commonly used approaches for the treatment of cancer.

Natural products have shown their potential role against various types of cancers, such as pancreatic, prostate, skin, gastric, lung, oral, blood, colorectal, liver, head and neck, cervical and breast cancers. Natural compounds may have bioavailability, efficacy, specificity, metabolism, and toxicity related issues when used as a drug. These therapeutic issues may be overcome by performing a series of necessary chemical changes in the lead compounds. A number of natural products have shown their therapeutic role against cancer in preclinical and clinical studies.



**Other Body Organs Affected** 

Fig. (1). Initiation, progression and metastasis of cancer.

Cancer is a uncontrolled division of cell due to the accumulation of defects *via* different mechanism, or mutations in DNA and the cells possess the capability to move from one part of body to another Fig. (1). Mutations or chromosomal aberrations affecting oncogenes and tumor suppressor genes causes malignant transformation of cells [3]. The most common sites of cancer are breast, cervix lungs, prostate, colorectal, stomach, and liver. Cancer cells can initiate, spread, and grow in various parts of body. The risk of developing cancer increases with age. Altering a diet that includes beneficial phytochemicals can have preventive and therapeutic role against cancer. In cancer chemoprevention, foods containing bioactive chemicals that have anticancer effect can be supplemented in diets. Alkaloids, flavonoids, terpenoids, polysaccharides, saponins, polyphenols and others have been reported as natural products with potential anticancer role [4]. Most of anticancer drugs that are in clinical use for cancer therapy originate from natural products derived from plants, marine sources, and microorganisms. Some natural products have shown their anticancer potential via regulating immune function, inducing apoptosis or autophagy, or inhibiting cell proliferation. Other

#### SUBJECT INDEX

#### A

Absorption 30, 156 glucose 30 ABTS 19, 27 assay 19 radicals scavenging assays 27 ACE 10, 15, 19, 25, 28, 29 activity 15, 19, 25 inhibited 29 substrates 28 ACEi 9, 10, 13, 14, 15, 18, 19, 24, 25, 29 activity 9, 10, 13, 14, 15, 18, 19, 24, 25, 29 fraction 25 peptides 29 ACE inhibitor(s) 9, 28, 29 peptides 9 ACE inhibitory (ACEi) 7, 8, 15, 19, 27, 28 activities 27 activity in vitro and antihypertensive activity 28 properties 28 Acetylcholinesterase 90 AChE 90 inhibition and antioxidant activity 90 inhibition and antioxidant activity combination 90 Acids 6, 15, 30, 32, 45, 48, 62, 67, 111, 126, 127, 136, 137, 139, 140, 143, 157, 159, 165, 168, 169 1,3,7-trimethyluric 139 3-butanoic 126 3-hydroxypilocarpic 140 3-Pyridylacetic 126 4-quinolone-3-carboxylic 48 5-hydroxyindoleacetic 137, 143 5-trimethoxybenzoic 136, 137 7-dimethyl uric 139 7-methyluric 139 acetic 6, 32 arachidonic 165 arylcarboxylic 67 betulinic 168, 169

butanoic 126

formic 32 lauric 30 methyluric 139 nitrous 67 oleanolic 157, 165 oxolinic 45 pipemidic 45 propionic 32 reserpic 136, 137 salvianolic 159, 168 trimethyluric 140, 143 ursolic 157, 165 Activation 3, 78, 154, 155, 163, 164, 165, 169, 172 carcinogenic 163 caspase-3 172 metabolic 78 of oncogenes 154, 155 Active 30, 131, 136 endolytic cysteine protease 30 human liver cytochrome enzyme 136 microsomal enzyme 131 Active metabolites 127, 130, 131, 133, 136, 137, 139, 144 identified 127 of dihydroquinidine 131 Activity 26, 27, 30, 44, 45, 46, 50, 51, 62, 63, 69, 70, 78, 81, 83, 89, 90, 124, 142, 143, 166, 168, 169, 170, 171 α-amylase 31 anticancer 170 antimicrobial 26, 30, 78 antimycobacterial 62 antiplasmodial 81 anti-thrombotic 30 antitumor 89 biodynamic 124 hTERT 166 motor 143 promising 90 spectrum 45 therapeutic 171 Adenomatous polyposis coli (APC) 155 ADMET 156, 173

Atta-ur-Rahman (Ed.) All rights reserved-© 2020 Bentham Science Publishers

Atta-ur-Rahman

of administered drug 156 properties 173 Agents 8, 154, 162, 163, 164, 165, 171 chemical 8 chemoprevention 163 chemopreventive 162, 163, 165 derived anticancer 171 dietary 162 marine-derived antitumor 163 microbial-derived chemotherapeutic 164 plant-derived 163 Agroindustrial byproducts 2, 5, 9, 18, 19, 24, 27, 28, 30, 31, 33 Alcalase and digestive enzymes 24 Alkaline 3, 7, 8, 9, 10, 11, 12, 13, 14, 15, 18, 31 extraction 3, 7, 8, 9, 10, 12, 13, 14, 15 extraction of proteins 8 protease 10, 11, 12, 14 serine-endopeptidase 18 Alkaloidal 124, 125 drug metabolism 125 drugs 124 Alkaloid drugs 125, 141, 144 important marketed 125 marketed 141 Alkaloids 124, 125, 126, 127, 130, 131, 133, 136, 139, 140, 144, 153, 169, 171 benzylisoquinoline 133 cholinergic agonist 125 cinchona 131 metabolism of 124, 144 plant 127 purine 139 tropane 127 vinca 169, 171 Alzheimer's disease 90 Amino acids 4, 6, 7, 15, 16, 17, 20, 27, 28, 30, 32, 50, 81, 86, 87 analysis 15, 16, 20 analyzer 20 composition, essential 20 decomposition 32 esters 87 -linker 86 sequences 4 aromatic 27, 28 essential 17, 32 Amylase 7, 110 impure food-grade 7

Amylopectin 108, 110, 111, 118, 120, 121 branching 108, 121 hydrolysis 120 natural 120 Anthraquinone derivatives 161 Antibacterial activity 49, 50, 54, 55, 59, 60, 64, 66, 70, 72, 73 Antibiotics 43, 44, 45, 51, 65, 77, 86, 87, 91, 93, 164 anti-tumor 164 broad-spectrum 43 functionalized quinolone 77 oxazolidinone class 65 prototype 4-quinolone 45 rifamycin class 91 Anticholinergic effects 126 Antifungal activities 60 Anti-hypertensive activities 31 Antihypertensive 25, 28, 29, 143 activity 28, 29, 143 drugs 28 peptides 25 Anti-infective chemotherapy field 43 Antimicrobial agents 30, 50, 73, 75, 82 designed new DNA-binding 75 natural 30 new potential 50 Antineoplastic activity 143 Antioxidant 1, 3, 7, 8, 18, 19, 27, 30, 31, 59 agents 59 capacity 27 generated 8 natural 27 Antioxidant activity 10, 11, 12, 14, 18, 19, 24, 27, 28, 33, 59, 64, 90 by ABTS 18 combination 90 highest 14 modulate 33 Antioxidant fractions 23, 25 active 25 purified 23 Antioxidant peptides 4, 15, 27 generating 4 Antithrombotic activity 24 Anti-tyrosinase 18, 19, 31 Anti-urease 59 APC activation 164 Apoptosis 25, 77, 151, 153, 155, 159, 162, 167, 168, 169, 170, 171, 172

#### Subject Index

increasing 162 inducing 25, 153 process 155, 169 Aristolochic acid 156, 157, 165 pathway 165 strong carcinogen 156 Aspergillus oryzae 4, 5, 30 Assays 5, 9, 15, 19, 20, 21, 28, 31, 92 bicinconinic 20 cell-based 31 first screening 28 radical scavenging 19 vitro biofilm 92 ATP 161, 168, 172 binding site 161, 168 Atropine 124, 126, 127, 128, 142 conjugated 128

#### B

Bacillolysin activity 6 Bacillus peptidases 4, 29 Bacillus 4, 17, 29, 30 licheniformis 4, 29, 30 pumilus 4 subtilis 4 suspensions 17 Bacterial type II topoisomerase enzymes 43 Behavior-modifying effects 142 Benzimidazole derivatives 75 Binding 45, 57, 60, 161, 166, 168, 172 ability 60 Bioactive compounds 2, 24, 50, 164, 166, 170 fractions 24, 164 molecule 170 Bioactive peptides 1, 3, 4, 5, 6, 8, 17, 23, 26, 29 generation of 1, 3 Bioactive properties of protein hydrolyzates 26 Biodegradable biopolymer 108 Biodynamic scaffolds 50, 52 Bioethanol industries 1 Biomass 2, 7, 18, 27, 108, 111, 120 extracting 7 **Biorefinery framework 32** Bituminaria bituminosa 160 Blood coagulation 31 Boltzmann's constant 116 Bovine serum albumin 20

#### Frontiers in Natural Product Chemistry, Vol. 6 185

Bradford assav 20, 21 BRCA2 genes suppress 155 Breast cancer 59, 152, 155, 165, 166, 168, 171 metastatic 171 positive 166 Brewer 7, 8, 10, 11, 12, 16, 20, 23, 24, 27, 28, 29.30.31 extracted 31 Broken rice by-product 3 Bromoenol lactone 165 Broussonetia papyrifera promoting autophagy 159 Byproduct 1, 3, 4, 6, 9, 10, 11, 12, 13, 14, 15, 17, 18, 19, 22, 23, 24, 27, 29 agri-food 6, 15, 22 corn 19 peptide sequence bioactivitya 24 potato 3, 18, 19, 27, 29 rice protein 17, 18, 19, 23

#### С

Callyspongia siphonella 162 Camellia sinensis 157, 158 Camptotheca acuminata 160 Cancer 151, 152, 153, 154, 155, 156, 162, 163, 164, 165, 167, 168, 169, 170, 171, 172.173 bladder 172 chemotherapeutic agents 172 chemotherapy 163, 168 colon 152, 155, 169 liver 152 metastatic 171 metastatic colorectal 168 neck 168 ovarian 172 pancreatic 152, 168 progression and metastasis of 153, 164 prostate 152, 165, 170 therapy 153, 156, 168, 171, 172, 173 thyroid 152 Cancer cells 14, 77, 153, 155, 162, 165, 166, 167, 168, 169, 170, 172 breast 166, 169, 172 growth 14 hematopoietic 165 pancreatic 166 proliferation 77 Capacity 14, 27, 30, 49, 86, 172

bile acid-binding 30 penetration 49 radical absorbance 27 radical scavenging 14 Capillary 22 electrophoresis 22 gel electrophoresis (CGE) 22 Capsicum 140 annum 140 frutescens 140 Carbohydrate hydrolysis 10 Carbohydrolases 1, 7, 8, 10, 11, 30, 31, 33 cocktails 8 gastrointestinal 30 use of 7 Carcinogenesis 154, 162 skin 77 Catharanthus roseus 136, 137 Cell death 167, 168, 169, 171, 172 programmed 167 Cell proliferation 153, 155, 170, 172 inhibiting 153 thyroid cancer 172 Cellulase 8.13 Cellulose 2 Characterization of protein hydrolyzates 19 Chemicals 152, 153 containing bioactive 153 toxic 152 Chemical structure 110, 127, 130, 132, 133, 138.139 of catuabine 130 of cytisine 127 of dihydroquinidine 133 of Dihydroquinine 132 of serotonin 139 of vinblastine 138 of vincristine 138 Chemopreventive agents isothiocyanates 162 Chinese herb 156 Chlorophyllin 162 Cholinergic transmission 90 Chromophore 26 Chymosin 5 Chymotrypsin 5 Chymotrypsina 5 Cognitive loss 90 Combination therapy of nutraceutical 170 Combinatorial peptide ligand libraries (CPLLs) 23

Corolase 5, 6, 10, 11, 12, 29, 30 pancreatic proteinase preparation 29 CPLLs technology 23 *Cribrochalina vasculum* 161 Cross-presentation antigen induction 164 Crude protein 19 *Curcuma longa* 158, 159 Curcumin downregulate 170 Cyclin 154, 161, 172 -CDK complexes 154 dependent kinases (CDKs) 154, 161, 172 Cyclodehydration 67 Cyclooxygenases 165

#### D

Deaths, cancer-related 170 Defatted wheat germ protein 9, 15 Degradation, metabolic 57 Dementia impressing 90 Demethylation 127, 139, 140 oxidative 127 process 139 Denaturation 15, 168 of expression of GFRs 168 Derivation of intrinsic viscosity 116 Derivatives 61, 67, 73, 79, 86, 133, 163, 172 2-aminothiazole 73 4-cyclic amino 79 biphenyl 172 chemical 163 phenanthrene 133 tricycle-fused quinoline-3-carboxylic acid 61 Design 70, 71, 90 AChE inhibitors 90 of oxazolidinone-ciprofloxacin hybrids 70 of oxazolidinone-levofloxacin hybrids 71 Detection 20, 23, 108, 110, 114, 125, 136, 152 differential refractive index 108 fluorescence 20, 23, 136 Detector 20, 22, 113, 114 diode-array 22 low scattering angle 114 photometric 20 Detector signal alignment 114 Dietary mineral bioavailability enhancement 26 Dietary supplements 164, 170, 171 natural 164

#### Atta-ur-Rahman

#### Subject Index

Differential scaning calorimetry 17 Digestive enzymes 24 Diseases 28, 29, 73, 84, 151, 152, 163, 170, 171 cardiovascular 29, 84 complex 151 coronary heart 28 curing 163 infective 73 renal 28 DLS 116, 121 analysis 116 measurement 121 DNA 45, 60, 69, 79, 153, 154, 155, 162, 163, 166.169 adduct formation 162 cleaved 169 enzyme-cleaved 169 new telomeric 166 repair mechanism 154, 155, 163 replication 69, 169 synthesis 45, 154 DNA gyrase and topoisomerase IV 43, 45, 47, 48.69.84 inhibition of 48, 69

#### Е

Effects 13, 162, 164, 169, 170, 171 antagonistic 170 antibacterial 13 anti-cancer 164, 169 anti-tumor 171 enhancing 162 Efficacy 43, 45, 50, 69, 152, 162, 170, 173 therapeutic 50, 170 Efflux pumps 44, 50, 51, 64, 85 bacterial 64 inhibitors (EPIs) 50, 51 up-regulated 51 Efflux pump system 50, 51 inhibitors 51 EGF-mediated cell migration 159, 169 Elemental analysis 19, 20, 21 **Emulsifying 31** Endoglucanase 8, 11 Enzymatic hydrolysis 1, 3, 4, 9, 15, 29, 127 microwave-assisted 15 Enzymatic 7, 12 hydrolysis by alkaline protease 12

methods 7 Enzyme(s) 4, 5, 6, 7, 8, 13, 14, 15, 19, 26, 27, 28, 29, 31, 32, 45, 139, 151, 152, 156, 162 activities 6 activity, main 7 -assisted extraction peanuts kernels 26 cytochrome P450 139 **DNA 45** liver 156 induction 162 inhibition 162 specificity 4 Epidermal growth factor receptors (EGFR) 159, 161, 168, 169 ER-independent mechanisms 166 ER-positive breast cancer 166 Estrogen receptor 158, 166 signaling 158, 166 Expression 155, 159, 162, 168, 169, 170, 172 gene 162 ligand-induced 159 suppressing GFRs 168 Extracellular 4, 168 domain 168 proteases 4 Extraction 2, 6, 7, 8, 10, 15, 17, 18, 22, 26, 110.127 alkaline-ethanol 18 aqueous 18 by chemical methods and hydrolysis 6 by enzymatic methods and hydrolysis 7 by physical/physical-chemical methods and hydrolysis 8 electro-assisted 8 enzyme-assisted 22 high pressure-assisted 8 of phenolic compounds and carbohydrate hydrolysis 10 oil 7 Extraction conditions 7, 15 subcritical water 15 Extraction methods 6, 15, 23, 136 liquid-liquid 136 Extracts 8, 151, 163, 164 crude 164 herbal 151

#### F

Ferric reducing antioxidant power (FRAP) 11, 12.15.27 Fine starches 108, 109, 111, 112, 113, 114, 116, 118, 119, 120, 121 solution 108, 109, 112, 113 structure of 108, 109 Fish by-product 15 Flavonoid-floxacin conjugates 85 Fluconazole 57, 60, 61 antifungal 60 Fluorescence 22, 26 detectors 22 spectroscopy 26 Fluoroquinolone 60, 70, 85, 87 class 60 conjugates 85 core 70 monomer 87 Food 3, 30, 31, 32, 33, 109 additives 31 enzymes 32 industry 3, 30, 109 ingredients 31, 32 practices 2 preservation 3, 30 preservatives 33 production 31 Formation 32, 48, 54, 57, 59, 67, 137, 170 biofilm 54 spindle 170 FQs-resistant pathogens 90 Fraction 10, 11, 14, 16, 28, 30, 31 active antimicrobial 30 protein-rich 31 solid 10, 11 Fruits 3, 8, 13, 14, 16, 84, 140, 157, 160, 170 oil-bearing 13, 16 seeds 3 Fungal infection 136 Fusarium oxysporum 159, 160

#### G

*Garcinia indica* 157 Gastrointestinal digestion 4, 10, 27, 29, 31 enzymes 4 simulated 10, 27, 29, 31 Gastrointestinal disorders 50 GC-MS techniques 133 Gel filtration chromatography 23, 30 Genes 153, 154, 155, 170 regulatory 155 tumor suppressor 153 tumour suppression 154 tumour suppressor 155 Genetic alternations 154 GFR genes 155 Glioblastoma 168 Grain 3, 6, 7, 8, 10, 11, 12, 16, 23, 24, 27, 28, 29.30.31 hydrolyzates 23, 31 peptides 31 proteins 29, 30 Green peppers 158 Growth 2, 30, 45, 152, 155, 165, 170 abnormal 152 bacterial 45 demographic 2 inhibitory activity 30 signalling pathways 155 tumor cells 170 Growth factor 155, 168 receptors (GFRs) 155, 168 receptors, epidermal 168 Gyrase 45, 50, 93

#### Η

Hematopoietic system 164 Hemicelluloses 2 Hepatic 124, 136, 144 metabolism accounts 136 microsomal enzymes 124, 144 Hepatotoxicity 164 HER2 phosphorylation 159, 169 Herbal medicines 156 High-performance liquid chromatography (HPLC) 20, 22, 23, 113, 125, 130, 136, 137, 144 Hodgkin's sickness activity 143 Homogenization and alkaline extraction 15 HPLC modalities 24 Human 137, 167 hepato-cellular carcinoma 167 liver cytochrome enzymes 137 Hybrids 59, 60, 63, 64, 66, 68, 69, 70, 71, 72, 73, 80, 83, 84, 85, 92

#### Atta-ur-Rahman

#### Subject Index

4-quinolone-triazole 63, 64 ciprofloxacin-pyrimidine 83 ciprofloxacin-triazole 60 flavonoid-fluoroquinolone 85 fluoroquinolone-azole-thiomorpholine 66 fluoroquinolone-triazole 59 metronidazole-quinolone 80 norfloxacin-thiazolidinone 72 oxadiazole-fluoroquinolone 68 oxadiazole-quinolone 68 quinolone-oxazolidinone 71 synthesized clinafloxacin-fluconazole 60 thiazole-quinolone 73 Hydrolysis 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 18, 19, 23, 26, 29, 108, 109, 118, 120, 121, 139 acid 109 casein 29 degree of 4, 9, 16, 18, 23 reaction fields 118 stereoselective 5 waxy rice starch 108, 120, 121 Hydrolyzates 3, 4, 9, 10, 15, 18, 23, 27, 28, 29.30.33 antioxidants 23 bioactive 4, 33 produced 29 seed 29 Hydrolyze membrane phospholipids 165 Hydrophilic interaction chromatography 24

#### Ι

Immobilized enzymolysis treatment 18 Immunodeficient patients 44 Immunomodulatory activity 8, 12, 25, 31 Industry 3, 25, 109, 120 brewing 3 rice starch 25 Infections 43, 44, 45, 71, 78, 87, 92 bacterial 44, 45 biofilm-associated 92 upper airway 44 urinary tract 45 Infectious diseases 44, 69, 91 persistent 91 Inflammatory cytokines 165 Inhibition 14, 15, 28, 29, 30, 44, 48, 50, 51, 75, 156, 157, 158, 159, 160, 161, 163, 166, 169, 171

Frontiers in Natural Product Chemistry, Vol. 6 189

α-amylase 30 EGFR phosphorylation 159 lipid peroxidation 14, 15 low ACE 29 of AKT and STAT3 growth inhibition 159 of EGF-mediated cell migration 159, 169 of estrogen receptor signaling 158, 166 of HER2 phosphorylation 159, 169 of lipoxygenases 157 of phospholipase 156, 157 of telomerase 158, 166 of topoisomerase 160, 161, 169 Inhibitory activity 10, 25, 30 angiotensin I-converting enzyme 25 greatest α-glucosidase 30

#### K

Kidney dysfunction 28 Kinase 161, 168, 172 receptor tyrosine 161, 168

#### L

Laburnum anagyroides 125 Light scattering peak intensity 114 Linker 68, 84, 88, 89, 172 -connected pyrimidine-quinolone hybrids 84 ethylene 88, 172 fluorophenylene 68 thiadiazole 89 Lipophilic substituents 48 Lipopolysaccharide-stimulated RAW 12 Liposome-based nanocarriers 168 Lipoxygenases 157, 165 inhibiting 165 Liquid chromatography 17, 20, 22, 125, 127, 133, 136, 141 high-performance 20, 125 -mass spectrometry 136, 140 normal-phase 136 normal-phase high-performance 136 rapid 133 sensitive 127 size-exclusion 17 Liquid delivery pump 113 Low-density lipoprotein cholesterol 30 Lung cancer 152, 165, 168 non-small-cell 168

Lung tumorigenesis, induced 162 Lysophospholipids 165

#### Μ

Magnolia officinalis 159 promoting 159 Major metabolites 132, 136, 137, 138, 141 of choline 141 of ergotamine 136, 137 of quinidine 132 of reserpine 136, 137 of yohimbine 136, 138 MAP kinase 155 pathways 155 Mass spectrometry (MS) 1, 22, 23, 26, 125, 133, 137, 141 chromatography-tandem 141 high-performance liquid chromatography/tandem 137 ion trap 133 liquid-chromatography 125 Mass spectrometry method 127 Matrix assisted laser desorption ionization (MALDI) 26 Mechanism 27, 32, 84, 151, 152, 153, 164, 165, 167, 168, 171, 172, 173 biochemical 173 dual-acting 84 therapeutic 164 Medicines 109, 144, 156, 162, 171 first herbal anti-cancer 171 traditional 162 traditional Chinese 162 Membrane-bound ectoenzyme 28 Metabolic enzymes of arachidonic acid pathway 165 Metabolic pathways 124, 126, 127, 130, 131, 132, 134, 135, 136, 140, 141, 144 for caffeine 140 for codeine 135 for ephedrine 135 of capsaicin 141 of pilocarpine 140 Metabolic syndrome 28, 29 cluster 28 Metabolism 30, 79, 124, 125, 127, 131, 133, 136, 137, 139, 140, 141, 152, 156, 162, 163.166 cellular 163

estrogen 166 glucose 30 modifying carcinogen 162 of capsaicin 140, 141 of cocaine 127 of vohimbine 136 Metabolism pathway 126, 128, 129, 134, 139 of atropine 128 of caffeine 139 of nicotine 126 of scopolamine 129 Metabolites 33, 124, 125, 126, 130, 133, 134, 136, 137, 139, 141, 142, 156, 166 conjugated 142 genotoxic 166 primary 125, 136 quinine 130 secondary 124 Metabolite Study 125, 126, 133, 136, 139, 140 of imidazole group of alkaloid 139 of indole group of alkaloids 136 of isoquinoline group of alkaloid 133 of phenanthrene group of alkaloid 133 of phenylethylamine group of alkaloids 133 of purine group of alkaloid 139 of pyridine group of alkaloids 125 of quinoline group of alkaloids 130 of terpenoid group of alkaloids 140 of tropane group of alkaloids 126 Methicillin-resistant Staphylococcus aureus (MRSA) 44 Methyl uric acid 140 Microbial 4, 8, 18, 27, 28 alkaline proteases 4 enzymes 4, 8, 18, 27, 28 Microorganisms 4, 44, 71, 92, 153 exogenous 4 resistant pathogenic 92 Microtubule 169, 172 dynamics, suppressing 169 polymerization 172 Modifications 48, 49, 50, 60, 74, 92, 151 chemical 48, 151 enzymatic 92 structural 47, 49, 50, 60, 92 Modulate transcriptional processes 172 Modulation 64, 157 of diverse transcription factors 164 of transcriptional factor 157 Molecular hybridization 43, 45, 64, 85

#### Atta-ur-Rahman

#### Subject Index

concept 64 technique 43 Molecules 50, 57, 59, 64, 86, 124, 144, 152, 156, 162, 164, 165, 170, 172 anti-apoptotic 165 antibiotic 86 cellular adhesion 165 chemopreventive 162, 165 complex 59 convenient chief 57 novel 64 roscovitine 172 signalling 156 therapeutic 152 Monoclonal antibodies 168 Mono-hydroxylation product 134 Mucosal signaling molecule 143 Mucuna pruriens 137 Multi-angle light scattering (MALS) 108, 109, 112, 113, 114 Mutation in GFR genes 155 Myc genes 155 Mycobacterium tuberculosis 92

#### Ν

Natural 30, 32, 54, 124, 151, 163, 166, 169 additives 32 bioactives 151, 166, 169 bioactive substance 151 drug discovery progam 163 electrophilic 54 medicines 124 preservatives 30 Natural product 163, 164 -based drugs 163 research 164 Neurodegenerative disorders 84 Neurological disorders 165 Neutral protease 10, 14 Neutrase 4, 13, 14, 18, 19, 28, 31 hydrolyzate 18 Nicotine 124, 125, 126, 142 Nitrogen-containing compounds 125 Norscopolamine 127, 129 conjugated 129 Nuclear magnetic resonance 141 Nutraceuticals 3, 28, 31, 170, 171 effective 170 promising 170

#### 0

Oil 2, 7, 8, 26, 116 extract 26 silicone 116 Olive 7, 13, 14, 16, 23, 28, 29 seeds 7, 23, 28, 29 stones 13, 14, 16 Oncogenes 153, 154, 155 viral 155 **Oncogenesis** 170 ORAC activity 27 Oral mucositis 164 Oxidation 139, 140, 141 process 139 reactions 139 Oxidative stress 90, 165, 168 Oxygen radical absorbance capacity (ORAC) 10, 11, 12, 15, 27

#### P

Palm kernel expeller proteins 13 Papain activity 6, 30 Parasympathetic nervous system 142 Pathogens 43, 44, 70, 71, 78, 87, 91, 152 anaerobic 78 gram-negative respiratory tract 70 intracellular 87 persistent 91 resistant 43, 71 Pathways 108, 120, 121, 133, 151, 155, 156, 165, 167, 168, 172 caspaseand non-caspase-dependent 172 mechanistic 167 mitochondrial 167 molecular 151 regulated 167 signalling 155 targeting multiple 165 Pectinase 7, 12, 13 Penicillium citrinum 160 Pepsin 5, 14 Peptidases 1, 2, 3, 4, 5, 6, 7, 8, 15, 17, 18, 27, 31 alkaline 7 dipeptidyl 5 microbial 8

staphylococcal 5 Peptidomics 2 Phenethyl isothiocyanate inhibition 162 Phenolic compounds 10, 26 Phenylethylamine group 133 Photosynthesis 108 Piperazine 50, 64, 66 moiety 66 Plant 4, 124 enzymes 4 metabolites 124 Plant peptides 1, 27 obtaining antioxidant 27 Plumbago zevlanica 161 Plum seeds 14, 16 Polyethylene glycol 87 Polyfluoroalkyl ketones 165 Polymers 109, 110, 112, 118, 164 branching amylopectin 110 natural 164 Polyphenols Cell division 151 Potency 48, 49, 57, 162, 172 cytotoxic 172 Potent antibacterial 57, 69, 77 agents 57 compounds 69 oxime 77 Pregnane X receptor (PXR) 162 Process 8, 109, 111, 113, 155, 156, 162, 168 homeostatic 168 multi-stage 162 resource-intensive 8 Processing 2, 32 sustainable 2 thermal 32 Production 2, 156 secondary metabolite 156 sustainable 2 Promoting apoptosis 158, 159 Promoting autophagy 159 chili peppers 159 Prostate 152, 153 Protease 11, 12, 14 Proteinase 5 Protein extraction 6, 7, 8, 15 process 6 Protein hydrolyzates 1, 3, 6, 19, 23, 24, 26, 27, 29, 30, 31, 32, 33 bioactive 3 obtaining 1, 3

Protein kinase C (PKC) 161 Protein quantification 22 Proteins 2, 3, 4, 5, 6, 8, 9, 10, 11, 14, 15, 16, 17, 18, 20, 21, 22, 23, 26, 28, 30, 31, 32, 92, 155, 165, 172 activator 165 anti-apoptotic 172 bacterial 92 canola 26, 28 cereals 6 denature 32 electro-activated 21 extracted 8 hydrolysis of 5, 6, 15, 30, 31 hydrolyzed 17 napin 26 precursor 3 rice residue 26 seed 9, 23 separation of 22, 23 signal-transducing 155 soluble 20, 22 technical 8 toxic 32 tumour suppressor 155 unhydrolyzed 16 wheat germ 3, 28 Protein synthesis 68, 69 inhibition 69 containing bacterial 68 inhibitor 69 Proteolysis 1, 7, 8, 9, 17, 18, 32, 33 Proteolytic enzymes 4 exogenous 4 Proteolytic system 4 Pump inhibitors 50, 51 bacterial efflux 50 Pyrazinamide derivatives 81

#### Q

Qubit assays 20 Quinolone 50, 63, 70, 74, 75, 81, 87, 89, 90, 92 acids 87 derivatives 74, 90 integration 69, 81 pharmacophores 70, 92 -pyrazol hybrids 75 skeleton 50, 63, 89

#### Atta-ur-Rahman

Subject Index

#### R

Radiotherapy 164 Rasemic form 70 Ras protein 155 Reactions 56, 59, 65, 65, 68, 109, 111, 133 cyclisation 66 mannich 59, 65 organic 59 Reactive 90, 109, 163, 166 carcinogenic metabolites 163 extrusion 109 metabolic intermediates 78 oxygen species 90, 166 Receptor 60, 143, 161, 168 growth factor-1 161 targeting tyrosine kinase 168 tyrosine kinase (RTK) 161, 168 Refractive index (RI) 110, 113, 114 differential 113 increment 114 Regulation 28, 31, 166 blood pressure 28 respective Novel Food 31 Regulatory proteins 154 Resistance 43, 44, 47, 48, 50, 69, 91, 92, 168 efflux-mediated 50 genetic material encoding antibiotic 43 rifamycin 91 Resistant Streptococcus pneumoniae 44 Reversed phase (RP) 16, 17, 23, 24 liquid chromatography 17 Rice 23, 110 genotypes 110 protein byproduct hydrolyzates 23 starch granules 110 Rice byproducts 4, 19, 27, 28, 30 broken 27 Rifampin 91, 92 -containing regimens 92 -quinolone hybridization 91 RNA polymerase 172

#### S

Salivary glands 143 Salvia 159 miltiorrhiza Promoting 159

officinalis Promoting 159 Scutellaria 157 baicalensis 157 lateriflora 157 SDS-polyacrylamide gel electrophoresis 21 Sequential extraction and hydrolysis (SeEH) 4, 6, 9, 15, 16 Serine endopeptidase 6 Serotonin 137, 139 metabolites 137 Sesamum indicum 158 Short interfering RNA (SRNA) 168 SiEH methods 7, 18, 33 direct 7, 33 Signals 151, 155, 168 anti-apoptotic 168 transmitting 155 Silybium marianum 159 Simultaneous extraction and hydrolysis (SiEH) 4, 15, 17, 18 SiSH methods 17, 32, 33 Six-membered heterocycles 50 Sodium dodecyl sulfate (SDS) 7, 17 -polyacrylamide gel electrophoresis 17 Solid-to-liquid ratio 15, 19 Spray-drying processes 110 Stabilize DNA strand breaks 45 Staphylococcus epidermis 44 Starch 3, 108, 109, 110, 111, 112, 115, 118, 120, 121 composition and chemical structure 110 concentration 109 conformation in solid state 108 fine waxy 110 granules 109, 111 nanoparticles 108, 109, 121 waxy barley 110 waxy corn 110 waxy rice 110, 118 Starch particles 109 nano-scale 109 Stokes-Einstein equation 116 Storage proteins 16 Structures 30, 44, 50, 51, 60, 62, 65, 67, 109, 110, 116, 133, 162 amphiphilic 30 carcinogen 162 crystalline 110 random coil 116 semi-crystalline 110

spherical 116 starch macromolecular 109 thiosemicarbazide 67 Suspensions 3, 4, 19, 27 complete cells 27 microbial 3 Sustainable circular bioeconomy model 2 Synergistic effects 156, 171 of compounds 171 Synthesis 43, 45, 47, 48, 50, 51, 52, 53, 54, 55, 60, 61, 62, 66, 72, 73, 74, 76, 77, 78, 79, 82, 85, 89, 90, 170 combinatorial 72, 170 mediated 66 of ciprofloxacin 53, 54 of fluoroquinolone-benzimidazole conjugates 76 of fluoroquinolones 89 of nalidixic acid hybrids 62 of Norfloxacin and Ciprofloxacin 52 of N-thiolated fluoroquinolones 55 of quinolones 82 of rosoxacin 51 Synthetic 43, 52, 57, 68, 85 organic chemistry 43 pathway 52, 57, 68, 85 Systems biology modelling 151

#### Т

Targeting 165, 166, 168, 169, 171 arachidonic acid pathway 165 microtubules 169, 171 receptors 168 telomerase 166 Tea 158, 159, 163, 166 black 163 green 158, 159, 163, 166 Telomerase 151, 155, 158, 166 human reverse transcriptase 166 Telomerase activity 155, 166 blocking 155 limited 166 Telomere length 166 Telomeres 155 Therapeutic 87, 125, 151, 152, 170 effect 87, 125, 151, 170 issues 152 Therapeutic role 151, 152, 153, 156, 166, 167, 170

of natural compounds 167 potential 170 Thermal treatments 6, 7, 18 applied 7 Thermolysin activity 6 Topoisomerase 64, 75, 77, 89, 93, 151, 160, 161, 169, 170 inhibitors 75, 170 Topoisomerase II 162, 169 inhibitor 162 Torsional stress 45 Toxicity 50, 124, 125, 152, 164 gastrointestinal 164 Toxic limitations 171 Traditional chinese medicine (TCM) 162 Transcription factors 151, 154, 155, 164, 165, 170 oncogenic 165 pro-inflammatory 164, 165 Transformation 141, 153, 155 malignant 141, 153 Transform-infrared spectroscopy 26 Trolox equivalent antioxidant capacity (TEAC) 14, 15, 27 Trypsin 5, 14 Tryptophan 17, 20 Tumour 155, 168 shrinkage 168 suppressor genes adenomatous polyposis coli 155 Tyrosine kinase inhibitors 168

Atta-ur-Rahman

#### U

Unripe capsules 133

#### V

Vancomycin-resistant Enterococci (VRE) 44

#### W

Waals force 60 Walnut 14, 25, 30 oil extraction residue 14, 25 residue 30 Water 2, 8, 10, 11, 12, 15, 18, 31, 50, 112, 114, 172 deionized 18

#### Subject Index

distilled 112 -bath, traditional 15 binding capacity 31 reduced 8 solubility 61, 172 -soluble form 172 Waxy rice starch 111, 120 granules 111 hydrolysis pathway 120 Ways of producing plant protein hydrolyzates 4

### Z

Zingiber officinale promoting 159



## PROF. DR. ATTA-UR-RAHMAN, FRS

Prof. Atta-ur-Rahman, Ph.D. in Organic Chemistry from Cambridge University (1968) has 1,232 international publications (45 international patents and 341 books). He received the following awards: Fellow Royal Society (FRS) London (2006), UNESCO Science Prize (1999), Honorary Life Fellow Kings College, Cambridge University (2007), Academician (Foreign Member) Chinese Academy of Sciences (2015), Highest Civil Award for Foreigners of China (Friendship Award, 2014), High Civil Award Austria ("Grosse Goldene Ehrenzeischen am Bande") (2007), Foreign Fellow Chinese Chemical Society (2013), Sc.D. Cambridge University (UK) (1987), TWAS (Italy) Prize (2009). He was the President of Network of Academies of Sciences of Islamic Countries (NASIC), Vice President TWAS (Italy), Foreign Fellow Korean Academy of Science & Technology, President Pakistan Academy of Sciences (2003-2006) and (2011 - 2014). He was the Federal Minister for Science and Technology of Pakistan (2000 – 2002), Federal Minister of Education (2002) and Chairman Higher Education Commission/Federal Minister (2002-2008), Coordinator General of COMSTECH (OIC Ministerial Committee) (1996-2012), and the Editor-in-Chief of Current Medicinal Chemistry.